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21. <input checked="" type="checkbox"/> The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492(a) (1) - (5)):			CALCULATIONS PTO USE ONLY
Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO.....			\$1040.00
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NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

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Karen Magri
Karen A. Magri, Registration No. 41,965
Date: March 29, 2002

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PATENT

In re: Application of Aubin et al.
Serial No.: To be Assigned
Filed: Concurrently Herewith
For: ESTROGEN RELATED RECEPTOR
ERR α , A REGULATOR OF BONE
FORMATION

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Sir:

In the Specification.

--Related Application Information

In the Claims.

9. (Amended) The method of Claim 1 wherein the mammal is a human.

In re: Application of Aubin et al.
Serial No.: To be Assigned
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Remarks

Claims 1-15 are pending in this application. Claim 9 has been amended herein to remove multiple dependencies from this claim. A marked up version of the amendment is attached hereto and is captioned "Version with Markings to Show Changes Made". It is submitted that this application is now in condition for substantive examination, which action is respectfully requested.

Respectfully submitted,



Karen A. Magri
Registration No. 41,965



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Version with Markings to Show Changes Made

9. (Amended) The method of [any of claims 1 to 8] Claim 1
wherein the mammal is a human.

**ESTROGEN RELATED RECEPTOR, ERR α , A REGULATOR OF BONE
FORMATION**

Field of the Invention

- 5 The present invention relates to methods and pharmaceutical compositions for modulation of bone formation.

Background of the Invention

- 10 In the description which follows, references are made to certain literature citations which are listed at the end of the specification and all of which are incorporated herein by reference.

- Nuclear receptors are transcription factors involved in various physiological regulatory processes. The superfamily to which nuclear receptors belong comprises both ligand-dependent molecules such as the steroid hormone-, thyroid hormone-, retinoic acid- and vitamin D-receptors, and an increasing number of so-called orphan receptors for which no ligand has yet been determined (Gronemeyer H, Laudet V., 1995; Enmark and Gustafsson, 1996). Indeed, it is not yet known whether the orphan receptors have ligands that await identification or whether they act in a constitutive manner. The orphan receptors display the same structural organization as do the classic ligand-dependent receptors: the A/B domain located in the N-terminal part of the protein harbors a ligand-independent transactivation function (AF-1); the C domain, which is the most conserved part of the molecule, is responsible for the specific DNA-binding activity; the E domain contains the ligand binding hydrophobic pocket and contributes to receptor dimerization and to the ligand-dependent transactivation function (AF-2).
- 15 20 25

- Two orphan receptors, estrogen receptor-related receptor α (ERR α) and ERR β (Giguere et al., 1988; NR3B1 and NR3B2, respectively, according to the Nuclear Receptors Nomenclature Committee, 1999) are closely related to the estrogen receptors ER α and ER β (Green et al., 1986; Kuiper et al., 1996; NR3A1 and NR3A2 respectively). ERR α (Genbank Accession No. for
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human $ERR\alpha$: NM_004451) and $ERR\beta$ were identified by low-stringency screening of cDNA libraries with a probe encompassing the DNA-binding domain of the human estrogen receptor (ER). Recently, a third estrogen receptor-related receptor, $ERR3$ or $ERR\gamma$, was identified by yeast two-hybrid screening with the glucocorticoid receptor interacting protein 1 (GRIP1) as bait (Hong et al, 1999). The DNA binding domain region of ERRs and ERs is highly conserved, however the others parts of the protein share very little homology (Giguere et al, 1988; Hong et al, 1999). Therefore, sequence alignment of $ERR\alpha$ and the ERs reveals a high similarity (68%) in the 66 amino acids of the DNA-binding domain and a moderate similarity (36%) in the ligand-binding E domain, which may explain the fact that $ERR\alpha$ does not bind estrogen. Although ligands for the ERRs have not been clearly identified, the pesticides chlordane and toxaphene have been suggested to be potential ligands for $ERR\alpha$ (Yang and Chen, 1999). $ERR\alpha$ has been identified as a regulator of the SV40 major late promoter during the early-to-late switch of expression (Wiley et al., 1993) and as a regulator of fat metabolism (Sladek et al., 1997; Vega et al, 1997). Yang et al. also showed that $ERR\alpha$ modulates the activating effect of estrogens on the lactoferrin promoter and suggested that $ERR\alpha$ may interact with ERs through protein-protein interaction (Yang et al., 1996; Zhang and Teng, 2000). Finally, $ERR\alpha$ has been described as a modulator of the human aromatase gene in breast, and hypothesized to be critical for normal breast development and to play an important role in the pathogenesis and maintenance of breast cancer via its ability to interact with ERs (Yang et al, 1998).

Postmenopausal osteoporosis is a condition caused primarily by the severe decrease of serum estrogen levels after cessation of ovarian function. The absence of estrogen results in an increase in bone turnover (Turner et al, 1994) and a negative bone remodeling balance, leading to bone loss and an increased fracture risk. An anabolic effect of estrogens on bone homeostasis has been documented in post-menopausal osteoporosis (for review see Pacifi, 1996), where bone loss can be reversed by administration of natural or

synthetic estrogens. Although the bone preserving effect of estrogen replacement is indisputable, the molecular and cellular mechanism of action for this hormone effect remain unclear. ERs are expressed in osteoblasts (Turner et al., 1994; Eriksen et al, 1988; Komm et al, 1988), and estrogens have been found to elicit effects ranging from modulation of gene expression to regulation of proliferation in this cell type (for review Harris et al, 1996). In contrast, mice lacking a functional ER α or ER β have only minor skeletal abnormalities (Korach et al, 1994; Windahl et al, 1999) suggesting that other mechanisms or receptors might be important during skeletal development.

ERR β expression is restricted to early development and to a few adult tissues (Giguere et al., 1988; Pettersson et al., 1996). In contrast, ERR α has a broader spectrum of expression, including fat, muscle, brain, testis and skin (Bonnelye et al, 1997b). Strikingly, ERR α is also highly expressed in the ossification zones of the mouse embryo (in long bones, vertebrae, ribs and skull), and is more widely distributed in osteoblast-like cells than is ER α (Bonnelye et al., 1997a). Moreover it has been shown that ERR α positively regulates the osteopontin gene (Vanacker et al, 1998), an extracellular matrix molecule secreted by osteoblasts and other cells and thought to play a role in bone remodelling among other functions (Denhardt and Noda, 1998).

Summary of the Invention

The inventors have demonstrated the involvement of ERR α in the modulation of bone formation in mammals. Up regulation of ERR α increased osteoblast differentiation from progenitor cells and also proliferation of progenitor cells. Down regulation of ERR α caused inhibition of bone formation, with reduction of osteoblast numbers and differentiation. ERR α was shown to be expressed also in osteocytes in both calvaria and long bones, indicating a role in skeletal maintenance.

In accordance with one embodiment of the present invention,

a method of increasing proliferation of osteoblasts in a mammal comprises administering to the mammal an effective amount of an agent selected from the group consisting of:

- 5 (a) an estrogen related receptor alpha ($ERR\alpha$) agonist;
- (b) a substantially purified $ERR\alpha$ protein; and
- (c) a nucleotide sequence encoding $ERR\alpha$ protein.
- (d) an agent which enhances expression of a gene encoding an $ERR\alpha$ protein.

10 In accordance with another embodiment of the present invention, a method of increasing differentiation of osteoblasts in a mammal comprises administering to the mammal an effective amount of an agent selected from the group consisting of:

- (a) an $ERR\alpha$ agonist;
- (b) a substantially purified $ERR\alpha$ protein; and
- 15 (c) a nucleotide sequence encoding $ERR\alpha$ protein.
- (d) an agent which enhances expression of a gene encoding an $ERR\alpha$ protein.

In accordance with another embodiment of the present invention, a method of reducing proliferation of osteoblasts in a mammal comprises administering to the mammal an effective amount of an agent selected from the group consisting of:

- (a) an $ERR\alpha$ antagonist;
- (b) a purified antibody which binds specifically to an $ERR\alpha$ protein;
- (c) an antisense nucleotide sequence complementary to and
- 25 capable of hybridizing to a nucleotide sequence encoding an $ERR\alpha$ protein; and
- (d) an agent which reduces expression of a gene encoding an $ERR\alpha$ protein.

In accordance with another embodiment of the present invention, a method of reducing differentiation of osteoblasts in a mammal comprises

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administering to the mammal an effective amount of an agent selected from the group consisting of:

- (a) an $ERR\alpha$ antagonist;
- (b) a purified antibody which binds specifically to an $ERR\alpha$ protein;
- 5 (c) an antisense nucleotide sequence complementary to and capable of hybridizing to a nucleotide sequence encoding an $ERR\alpha$ protein; and
- (d) an agent which reduces expression of a gene encoding an $ERR\alpha$ protein.

10 In accordance with another embodiment of the present invention, a method for treating a disorder associated with bone loss in a mammal comprises administering to the mammal an effective amount of an agent selected from the group consisting of:

- (a) an $ERR\alpha$ agonist;
- 15 (b) a substantially purified $ERR\alpha$ protein; and
- (c) a nucleotide sequence encoding $ERR\alpha$ protein.
- (d) an agent which enhances expression of a gene encoding an $ERR\alpha$ protein.

20 In accordance with another embodiment of the present invention, a method for treating a disorder associated with unwanted bone formation comprises administering to the mammal an effective amount of an agent selected from the group consisting of:

- (a) an $ERR\alpha$ antagonist;
- (b) a purified antibody which binds specifically to an $ERR\alpha$ protein;
- 25 (c) an antisense nucleotide sequence complementary to and capable of hybridizing to a nucleotide sequence encoding an $ERR\alpha$ protein; and
- (d) an agent which reduces expression of a gene encoding an $ERR\alpha$ protein.

In accordance with another embodiment of the present invention, a method for screening a candidate compound for its ability to modulate $ERR\alpha$ activity comprises:

- 5 (a) providing a system for measuring a biological activity of $ERR\alpha$; and
- (b) measuring the biological activity of $ERR\alpha$ in the presence or absence of the candidate compound,

wherein a change in $ERR\alpha$ activity in the presence of the compound relative to $ERR\alpha$ activity in the absence of the compound indicates an ability to modulate $ERR\alpha$ activity.

In accordance with another embodiment of the present invention, a method for screening a candidate compound for potential efficacy in promoting bone formation comprises:

- 15 (a) providing an assay system for determining $ERR\alpha$ agonist activity of a compound; and
- (b) testing the candidate compound for $ERR\alpha$ agonist activity in the assay wherein $ERR\alpha$ agonist activity in the candidate compound indicates potential efficacy as a promoter of bone formation.

In accordance with another embodiment of the present invention, a method for screening a candidate compound for potential efficacy in inhibiting bone formation comprises:

- (a) providing an assay system for determining $ERR\alpha$ antagonist activity of a compound; and
- 25 (b) testing the candidate compound for $ERR\alpha$ antagonist activity in the assay wherein $ERR\alpha$ antagonist activity in the candidate compound indicates potential efficacy as an inhibitor of bone formation.

In accordance with another embodiment of the present invention, a pharmaceutical composition comprises an effective amount of an agent selected from the group consisting of:

- 30 (a) an $ERR\alpha$ agonist;

- (b) a substantially purified $ERR\alpha$ protein; and
- (c) a nucleotide sequence encoding $ERR\alpha$ protein and a pharmaceutically acceptable carrier.
- (d) an agent which enhances expression of a gene encoding an $ERR\alpha$ protein.

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In accordance with another embodiment of the present invention, a pharmaceutical composition comprises an effective amount of an agent selected from the group consisting of:

- (a) an $ERR\alpha$ antagonist;
- (b) a purified antibody which binds specifically to $ERR\alpha$ protein;
- (c) an antisense nucleotide sequence complementary to and capable of hybridizing to a nucleotide sequence encoding $ERR\alpha$ protein; and
- (d) an agent which reduces expression of the gene encoding $ERR\alpha$ protein

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and a pharmaceutically acceptable carrier.

Summary of the Drawings

Certain embodiments of the invention are described, reference being made to the accompanying drawings, wherein:

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Figure 1, Panel A is a Northern blot showing expression of $ERR\alpha$, alkaline phosphatase (ALP), osteopontin (OPN) and osteocalcin (OCN), in primary rat calvaria (RC) cells over a proliferation-differentiation time course in presence (+Dex) or absence (-Dex) of dexamethasone (Dex) during proliferation (day 6), early nodule formation (day 10) and nodule mineralization (day 15).

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Figure 1, Panel B shows $ERR\alpha$ mRNA expression normalized against that of the ribosomal protein L32; the Y-axis is the ratio of the $ERR\alpha$ signal to that of L32. For comparison, mRNA levels for three osteoblast markers, alkaline phosphatase (ALP), osteopontin (OPN) and osteocalcin (OCN), are also shown (Panel A) and normalized against L32 (Panel B).

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Figure 2 shows detection of mERR α , (Type I collagen, osteopontin, alkaline phosphatase, bone sialoprotein and osteocalcin) by RT-PCR in libraries selected based on the basic molecular phenotype of the poly(A) PCR libraries made from discrete isolated colonies at different stages of osteoblast differentiation and bone development. Gene expression profiles of colonies were determined by analyzing expression of several known osteoblast lineage markers. These 19 libraries, from a pool of >100 characterized libraries, were selected for fingerprinting on the basis that they represent several transitional stages: primitive progenitors (A), progressively more mature precursors (B,C,D) and terminally differentiated, bone forming osteoblasts (E). While category order is progressive, the order of colonies within each category is random.

Figure 3 shows in Panel A: a Western blot of whole-cell extracts obtained from Hela cells and, in Panel B: MC3T3-E1 transfected with the empty expression vector PcDNA3 and PcDNA3m ERR α , separated by SDS-PAGE (10% polyacrylamide). Panels C to M show immunolabelling for ERR α in RC cells over a proliferation-differentiation time course: during proliferation at day 2 (C), confluence (D), nascent nodules (E), mineralized nodules (F), osteoblasts (G). Negative control for anti-rabbit antibody is shown in (H). Immunolabelling for alkaline phosphatase ALP (I), bone sialoprotein BSP (J), osteocalcin OCN (K) and osteopontin OPN (L), are also shown. Negative control for anti-mouse antibody is shown (M).

Figure 4, Panels A to C, show photographs of 21 days fetal calvaria sections immunolabelled for ERR α . ERR α detection is seen in osteogenic front (A), more mature growing bone trabecula (B) and remodeling bone (C). Panel D shows immunolabelling for ERR α in adult calvaria, osteocytes (D). Panels F to I show immunolabelling for alkaline phosphatase ALP (F), bone sialoprotein BSP (G), osteocalcin OCN (I) and osteopontin OPN (H). Negative controls for anti-rabbit antibody and anti-mouse antibody are shown (E,J respectively).

Figure 5, panel A shows proliferation of RC cells treated with antisense/sense oligonucleotide at 0.5 μ M, 1 μ M and 2 μ M (AS or S) or no oligonucleotide (Ct) during the proliferation stage between days 1-6. Inhibition of ERR α protein synthesis inhibited the cell proliferation based on the cell number. Three 24 wells per treatment group were trypsinized and cells were counted. Data are expressed as the cell number mean \pm SEM and are representative of three independent experiment. ANOVA revealed a very highly significant ($p < 0.0001$) effect of antisense on cell proliferation. Panel B shows expression of markers of osteoblast differentiation. Total RNA was extracted and RT-PCR performed on triplicate samples using specific primers for early markers (ALP, BSP, OPN, cbfa1, COLLI), proliferation (Cyclin D1, c-Fos) and apoptosis (Bcl2, Bax) at day 6. Panel C shows PCR product normalized to L32 PCR product. ANOVA revealed a significant ($p < 0.05$) and a highly significant ($p < 0.001$) effect of antisense treatment for Cbfa1 and BSP respectively. **= $p < 0.01$, ***= $p < 0.001$ vs control (Student's unpaired *t*-test).

Figure 6, Panel A shows nodule formation in RC cells treated with antisense/sense oligonucleotide at 0.5 μ M, 1 μ M and 2 μ M or no oligonucleotide (Ct) during the proliferation stage between days 1-6 and then switched to normal differentiation medium. Inhibition of ERR α protein synthesis induced a decrease in bone nodules formation. Three 24 wells per treatment group were von Kossa stained and the nodules were counted. Data are expressed as the nodule mean \pm SEM and are representative of two independent experiment. ANOVA revealed a very highly significant ($p < 0.0008$) effect of antisense, on bone nodules formation. Panel B shows expression of osteoblast markers. Total RNA was extracted and RT-PCR performed on triplicate samples using specific primers for markers of osteoblast ALP, BSP, OPN, OCN, COLLI at day 15. Panel C shows PCR product normalized to L32 PCR product (C). ANOVA revealed a highly significant ($p < 0.001$) effect of antisense treatment for ALP and BSP respectively. *= $p < 0.05$, **= $p < 0.01$, vs control (Student's unpaired *t*-test).

Figure 7, Panel A shows a Northern blot of primary RC cells transfected at 50% of confluence using a pcDNA3 empty plasmid as a control

and pcDNA3-ERR α at 0.5 μ g of total DNA per transfection. As control, of efficiency of the transfection, total RNA of each group was extracted 72h after transfection and northern blot was performed with samples (pool from three 35-mm culture). Panel B shows nodule numbers in five 35-mm dishes per treatment group for three independent experiments, von Kossa stained and the nodules counted. ANOVA revealed a significant ($p < 0.01$) effect of overexpression of ERR α , on bone nodules formation. Panel C shows expression pattern, using specific probes for markers of osteoblast ALP, BSP, OPN, OCN, COLLI total RNA was extracted at 72h after transfection, day 10 and day 13, and northern were performed (pool from three 35-mm culture). Data were pooled from three independent experiments and the pattern of expression are presented.

Figure 8, Panel A shows expression of ERR α , ER α and ER β in primary rat calvaria (RC) cells over a proliferation-differentiation time course by RT-PCR in presence (+Dex) or absence (-Dex) of dexamethasone (Dex) during proliferation (day 6), early nodule formation (day 10) and nodule mineralization (day 15). Total RNA was extracted and RT-PCR performed using specific primers for ERR α , ER α and ER β . In Panel B, PCR product was normalized to L32 PCR product.

Figure 9: Expression of ERR α , ER α and ER β in primary rat bone marrow primary culture (RBM) cells over a proliferation-differentiation time course by RT-PCR in presence (+Dex) or absence (-Dex) of dexamethasone (Dex) during proliferation (day 4-6), early nodule formation (day 9-11) and nodule mineralization (day 14-17) (A). Total RNA was extracted and RT-PCR performed using specific primers for ERR α , ER α and ER β (A). PCR product was normalized to L32 PCR product (B).

Figure 10 shows expression of ERR α after normalization with L32 in primary rat bone marrow primary culture (RBM) cells over a proliferation-differentiation time course by Northern blotting in presence of dexamethasone (Dex 10^{-8} M), estrogen (E2; 10^{-8} M), or vitamin D₃ (1.250H D₃; 10^{-9} M) (Panel A), or PGE2 (10^{-9} M), TGF β (10^{-10} M) (Panel B) or 1-34 PTH (10^{-11} M)

(Panel C) during proliferation (day 6), early nodule formation (day 10-12) and nodule mineralization (day 15-16). An acute exposure of RC cells for 24 hours at beginning from day 9 (nascent nodule formation) or day 15 (mature nodules present) to either Dex, E2, D3 (A), PGE2, TGF β (B) or 1-34 PTH (C) were also shown.

Figure 11 shows effect of estrogen *in vivo* on ERR α expression level in mice. Mice were treated once weekly by subcutaneous injection with either vehicle (0.2ml corn oil) or 500 μ g of 17 β -estradiol. Total RNA from femur (Panel A) or flushed femur (Panel B) was extracted and RT-PCR performed using specific primers for ERR α . ERR α PCR product was normalized to L32 PCR in femur and flushed femur.

Figure 12 shows expression of ERR α in a rat model of postmenopausal osteoporosis. Female rats were either ovariectomized (OVX) or sham-operated (Sham). ERR α expression is increased 4 weeks post-surgery in the long bones of OVX, (B, C) versus Sham-operated (A) femurs, but not in calvaria bones (E, OVX vs F, Sham). Active osteoblasts and osteocytes (arrows, A, B) are intensely labeled as are osteoclasts (arrows, C) in sections from the OVX animals. Negative anti-rabbit antibody control is also shown (D).

Detailed Description of the Invention

The present invention demonstrates a new role for the orphan receptor, estrogen related receptor alpha (ERR α), namely the modulation of bone formation in mammals.

Bone formation occurs during fetal development and post-natal growth and also during adult life either at a low rate as part of normal bone remodelling or at an accelerated rate in response to injury or abnormal bone loss. Bone formation involves a number of processes, including osteoblast progenitor cell proliferation, osteoblast differentiation from progenitor cells and mineralisation of matrix produced by the osteoblasts. The inventors have shown that ERR α plays a role in all of these processes.

In a rat calvaria cell culture system, which is an accepted model of bone formation, it has been shown that upregulation of $ERR\alpha$ levels increased osteoblast differentiation and bone formation, while down regulation of $ERR\alpha$ led to inhibition of bone formation, with reduction of osteoblast numbers and differentiation, and a proliferation-independent, complete inhibition of both mineralised and unmineralised bone nodule formation.

$ERR\alpha$ has been shown in this system and in adult rat bone marrow stromal cell cultures, a second well-characterized system of osteoprogenitor cell proliferation and differentiation, to be expressed throughout all stages of osteoblastic differentiation. $ERR\alpha$ is more highly expressed in cuboidal osteoblastic cells than in surrounding non-nodular/fibroblastic cells, and nuclear expression of $ERR\alpha$ increased as osteoblasts matured. Immunocytochemistry showed that $ERR\alpha$ is also highly expressed in vivo in developing fetal rat calvaria, both in sutural cells and cells at the osteogenic front, on trabecular and remodeling bone. $ERR\alpha$ is also highly expressed in fetal and adult osteocytes in calvaria and in other bones including femurs.

A number of disorders are associated with bone loss or bone degeneration. Such disorders include osteoporosis, osteoarthritis, Paget's disease, periodontal disease, osteolytic bone tumour metastases in, for example, breast cancer and multiple myeloma, osteochondrodysplasias, osteogenesis imperfecta, sclerosing bone displasias and osteomalacia.

The present invention provides methods and pharmaceutical compositions for treating such disorders to promote bone formation, by increasing $ERR\alpha$ activity. $ERR\alpha$ activity may be increased in a subject either by increasing the amount of $ERR\alpha$ protein present or by stimulating the activity of existing $ERR\alpha$ protein. Increased $ERR\alpha$ activity may be achieved, for example, by up regulating expression of the $ERR\alpha$ gene, by gene therapy to provide a nucleotide sequence encoding $ERR\alpha$ protein, by administering an agent which enhances $ERR\alpha$ expression, by administering $ERR\alpha$ protein or by administering an $ERR\alpha$ agonist.

Another group of diseases involves unwanted or inappropriate bone formation. Such diseases include fibrodysplasia ossificans progressiva, osteoblastic bone metastases such as prostate cancer and osteosarcomas. The present invention provides methods and pharmaceutical compositions for treating such disorders by reducing $ERR\alpha$ activity. $ERR\alpha$ activity may be reduced by reducing the amount of $ERR\alpha$ protein being produced or by inhibiting the activity of $ERR\alpha$ protein. This may be achieved, for example, by administering an antisense sequence or an agent which reduces $ERR\alpha$ expression, an antibody which binds specifically to $ERR\alpha$ protein or an $ERR\alpha$ antagonist.

The invention also provides a method for screening a candidate compound for its ability to modulate $ERR\alpha$ activity in a suitable system, in the presence or absence of the candidate compound. A change in $ERR\alpha$ activity in the presence of the compound relative to $ERR\alpha$ activity in the absence of the compound indicates that the compound modulates $ERR\alpha$ activity. If $ERR\alpha$ activity is increased relative to the control in the presence of the compound, the compound is an $ERR\alpha$ agonist. Conversely, if $ERR\alpha$ activity is decreased in the presence of the compound, the compound is an $ERR\alpha$ antagonist.

Suitable systems for measuring $ERR\alpha$ activity include examination of osteoblast proliferation or osteoblast differentiation in rat calvaria cell cultures or in bone marrow stromal cell cultures as described herein or other systems known to those of ordinary skill in the art, such as organ cultures of calvaria or femur bones or injection over the calvaria in vivo.

In accordance with a further embodiment of the invention, the $ERR\alpha$ signalling pathway may be modulated by modulating the binding of the $ERR\alpha$ to an $ERR\alpha$ binding partner. Such a binding partner may include for example the estrogen receptor. $ERR\alpha$ can be used to upregulate the transcription and thus expression of genes which work together with $ERR\alpha$ to affect skeletal development.

The invention further provides methods for screening candidate compounds to identify those able to modulate signaling by $ERR\alpha$ through a pathway involving $ERR\alpha$.

For example, the invention provides screening methods for compounds
5 able to bind to $ERR\alpha$ which are therefore candidates for modifying the activity of $ERR\alpha$. Various suitable screening methods are known to those in the art, including immobilization of $ERR\alpha$ on a substrate and exposure of the bound $ERR\alpha$ to candidate compounds, followed by elution of compounds which have bound to the $ERR\alpha$.

10 The invention also provides a method of modulating a $ERR\alpha$ signaling pathway by increasing or decreasing the availability of $ERR\alpha$ or by modulating the function of the $ERR\alpha$.

The invention further provides methods for preventing or treating diseases characterised by an abnormality in an $ERR\alpha$ signaling pathway
15 which involves $ERR\alpha$, by modulating signaling in the pathway.

According to another aspect of the present invention is a method for suppressing in a mammal, the proliferation of a cell capable of being stimulated to proliferate by $ERR\alpha$, the method comprising administering to the mammal an effective amount of a $ERR\alpha$ antagonist or an antibody which
20 binds specifically to $ERR\alpha$. Such cells include but are not limited to primitive osteoprogenitor cells.

The invention also enables transgenic non-human animal models, which may be used for study of the effects of over and under expression of the $ERR\alpha$ gene, for the screening of candidate compounds as potential
25 agonists or antagonists of this receptor and for the evaluation of potential therapeutic interventions.

The transgenic animals of the invention may also provide models of disease conditions associated with abnormalities of $ERR\alpha$ expression. Animal species suitable for use in the animal models of the invention include
30 mice, rats, rabbits, dogs, cats, goats, sheep, pigs and non-human primates.

Animal models may be produced which over-express $ERR\alpha$ by inserting a nucleic acid sequence encoding $ERR\alpha$ into a germ line cell or a stem cell under control of suitable promoters, using conventional techniques such as oocyte microinjection or transfection or microinjection into stem cells.

5 Animal models can also be produced by homologous recombination to create artificially mutant sequences (knock-in targeting of the $ERR\alpha$ gene) or loss of function mutations (knock-out targeting of the $ERR\alpha$ gene). For example, knock-out targeting of the $ERR\alpha$ gene). For example, knock-out animal models can be made using the tet-receptor system described U.S. Patent No.

10 5,654,168 or the Cre-Lox system described, for example, in U.S.P. Nos. 4,959,717 and 5,801,030.

In accordance with one embodiment of the invention, transgenic animals are generated by the introduction of a $ERR\alpha$ transgene into a fertilized animal oocyte, with subsequent growth of the embryo to birth as a

15 live animal. The $ERR\alpha$ transgene is a transcription unit which directs the expression of $ERR\alpha$ gene in eukaryotic cells. To create the transgene, $ERR\alpha$ gene is ligated with an eukaryotic expression module. The basic eukaryotic expression module contains a promoter element to mediate transcription of $ERR\alpha$ sequences and signals required for efficient for termination and

20 polyadenylation of the transcript. Additional elements of the module may include enhancers which stimulate transcription of $ERR\alpha$ sequences. The most frequently utilized termination and polyadenylation signals are those derived from SV40. The choice of promoter and enhancer elements to be incorporated into the $ERR\alpha$ transgene is determined by the cell types in which

25 $ERR\alpha$ gene is to be expressed. To achieve expression in a broad range of cells, promoter and enhancer elements derived from viruses may be utilized, such as the herpes simplex virus thymidine kinase promoter and polyoma enhancer. To achieve exclusive expression in a particular cell type, specific promoter and enhancer elements could be used, such as the promoter of the

30 mb-1 gene and the intronic enhancer of the immunoglobulin heavy chain

gene. In a preferred embodiment, a bone specific promoter such as the bone sialoprotein promoter may be used to target expression in osteoblasts.

The $ERR\alpha$ transgene is inserted into a plasmid vector, such as pBR322 for amplification. The entire $ERR\alpha$ transgene is then released from the plasmid by enzyme digestion, purified and injected into an oocyte. The oocyte is subsequently implanted into a pseudopregnant female animal. Southern blot analysis or other approaches are used to determine the genotype of the founder animals and animals generated in the subsequent backcross and intercross.

Such deficient mice will provide a model for study of the role of $ERR\alpha$ in bone cell differentiation and proliferation and general skeletal development. Such animals will also provide tools for screening candidate compounds for their interaction with $ERR\alpha$ or the signalling pathway activated by $ERR\alpha$.

The invention also provides pharmaceutical compositions for promoting bone formation, comprising as active ingredient a substantially purified $ERR\alpha$ protein, an $ERR\alpha$ agonist or an isolated nucleotide sequence encoding $ERR\alpha$ protein.

Such compositions are useful, for example, in treating disorders associated with bone loss.

$ERR\alpha$ protein may be produced by conventional recombinant techniques permitting expression of $ERR\alpha$ by a suitable host cell. A DNA encoding $ERR\alpha$ may be prepared as described, for example, in Giguere et al. (1998). Techniques for production of proteins by recombinant expression are well known to those in the art and are described, for example, in Sambrook et al. (1989) or latest edition thereof. Suitable host cells include *E. coli* or other bacterial cells, yeast, fungi, insect cells or mammalian cells.

The invention provides for compositions for promoting bone formation comprising as active ingredient an $ERR\alpha$ agonist obtained by using a screening method as described herein.

It may be advantageous, in treating disorders associated with bone loss, to employ a combination therapy, first administering an agent such as a

biphosphonate to suppress osteoclast function, followed after a suitable period of time, by administration of a pharmaceutical composition for promoting bone formation, as described herein. Such treatment regimens are well known to those of ordinary skill in the art.

5 A nucleotide sequence encoding $ERR\alpha$ protein may be administered to a subject experiencing bone loss due to an absent or defective $ERR\alpha$ gene either in vivo or ex vivo. Expression may be targeted to a selected cell or tissue by use of an appropriate promoter, for example the bone-specific promoter for bone sialoprotein (Stein et al. (2000)). For example, stem cells
10 or bone marrow stromal cells may be obtained from a subject and treated in vivo with the nucleotide sequence, the cells then being restored to the subject. Such methods are described in Horwitz et al. (1999).

The invention also provides pharmaceutical compositions for reducing bone formation, comprising as active ingredient an antibody which binds
15 specifically to $ERR\alpha$, an $ERR\alpha$ antagonist or a negative regulator such as an antisense nucleic acid or a dominant negative mutant version of the $ERR\alpha$ gene.

The invention provides for compositions for reducing bone formation comprising as active ingredient an $ERR\alpha$ antagonist obtained by using a
20 screening method as described herein.

Antibodies which bind specifically to $ERR\alpha$ protein may be made by conventional techniques.

The term "antibodies" includes polyclonal antibodies, monoclonal antibodies, single chain antibodies and fragments such as Fab fragments.

25 In order to prepare polyclonal antibodies, fusion proteins containing defined portions or all of an $ERR\alpha$ protein can be synthesized in bacteria by expression of the corresponding DNA sequences, as described above. Fusion proteins are commonly used as a source of antigen for producing antibodies. Alternatively, the protein may be isolated and purified from the
30 recombinant expression culture and used as source of antigen. Either the

entire protein or fragments thereof can be used as a source of antigen to produce antibodies.

The purified protein is mixed with Freund's adjuvant and injected into rabbits or other appropriate laboratory animals. Following booster injections at weekly intervals, the animals are then bled and the serum isolated. The serum may be used directly or purified by various methods including affinity chromatography to give polyclonal antibodies.

Monoclonal anti-ERR α antibodies may be produced by methods well known in the art. Briefly, the purified protein or fragment thereof is injected in Freund's adjuvant into mice over a suitable period of time, spleen cells are harvested and these are fused with a permanently growing myeloma partner and the resultant hybridomas are screened to identify cells producing the desired antibody. Suitable methods for antibody preparation may be found in standard texts such as Barreback, E.D. (1995).

The pharmaceutical compositions of the invention may comprise, in addition to the active ingredient, one or more pharmaceutically acceptable carriers.

Administration of an effective amount of a pharmaceutical composition of the present invention means an amount effective, at dosages and for periods of time necessary to achieve the desired result. This may also vary according to factors such as the disease state, age, sex, and weight of the subject, and the ability of the composition to elicit a desired response in the subject. Dosage regima may be adjusted to provide the optimum therapeutic response. For example, several divided doses may be administered daily or the dose may be proportionally reduced as indicated by the exigencies of the therapeutic situation.

By pharmaceutically acceptable carrier as used herein is meant one or more compatible solid or liquid delivery systems. Some examples of pharmaceutically acceptable carriers are sugars, starches, cellulose and its derivatives, powdered tragacanth, malt, gelatin, collagen, talc, stearic acids, magnesium stearate, calcium sulfate, vegetable oils, polyols, agar, alginic acids, pyrogen-free water, isotonic saline, phosphate buffer, and other

suitable non-toxic substances used in pharmaceutical formulations. Other excipients such as wetting agents and lubricants, tableting agents, stabilizers, anti-oxidants and preservatives are also contemplated.

The compositions described herein can be prepared by known
5 methods for the preparation of pharmaceutically acceptable compositions which can be administered to subjects, such that an effective quantity of the active substance is combined in a mixture with a pharmaceutically acceptable carrier. Suitable carriers and formulations adapted for particular modes of administration are described, for example, in Remington's Pharmaceutical
10 Sciences (Remington's Pharmaceutical Sciences, Mack Publishing Company, Easton, Pa., USA 1985). On this basis the compositions include, albeit not exclusively, solutions of the substance in association with one or more pharmaceutically acceptable vehicles or diluents, and contained in buffered solutions with a suitable pH and iso-osmotic with the physiological fluids.

15 The pharmaceutical compositions of the invention may be administered therapeutically by various routes such as by injection or by oral, nasal, buccal, rectal, vaginal, transdermal or ocular routes in a variety of formulations, as is known to those skilled in the art.

The present invention also enables the analysis of factors affecting the
20 expression of the $ERR\alpha$ gene in humans or in animal models. The invention further provides a system for screening candidate compounds for their ability to turn on or turn off expression of the $ERR\alpha$ gene and the identification of binding partners which may also affect expression of $ERR\alpha$ or certain downstream partners.

25 For example, an RC cell culture system can be used to identify compounds which activate production of $ERR\alpha$ or, once $ERR\alpha$ production has been activated in the cells, they can be used to identify compounds which lead to suppression or switching off of $ERR\alpha$ production. Alternatively, such a cell culture system can be used to identify compounds or binding partners of
30 $ERR\alpha$ which increase its expression. Compounds thus identified are useful as therapeutics in conditions where $ERR\alpha$ production is deficient or excessive.

The present invention enables also a screening method for compounds of therapeutic utility as antagonists of the biological activity of $ERR\alpha$. Such antagonist compounds are useful, for example, to reduce or prevent differentiation and maturation of osteoblasts and osteocytes. $ERR\alpha$

5 antagonists may also be used in the treatment of bone related disorders involved in inappropriate bone cell growth. Those skilled in the art will be able to devise a number of possible screening methods for screening candidate compounds for $ERR\alpha$ antagonism.

A screening method may also be based on binding to the $ERR\alpha$ receptor. Such competitive binding assays are well known to those skilled in
10 the art. Once binding has been established for a particular compound, a biological activity assay is employed to determine agonist or antagonist potential.

15 **$ERR\alpha$ is expressed in osteoblast lineage cells throughout their developmental lifetime and $ERR\alpha$ mRNA and protein are more highly and widely expressed than either $ER\alpha$ and $ER\beta$.**

The inventors have shown that $ERR\alpha$ mRNA is expressed in differentiating primary cultures of RC cells and rat bone marrow stromal
20 (RBM) cells and in single isolated osteoblast colonies at all detectable stages of differentiation, suggesting that $ERR\alpha$ may have a function in osteoblasts throughout their developmental lifetime. $ERR\alpha$ mRNA is present at much higher levels than either $ER\alpha$ or $ER\beta$ mRNA in both RC and RBM cell cultures. $ERR\alpha$ protein was also found more widely distributed in vitro in RC
25 cell cultures than either $ER\alpha$ or $ER\beta$. $ERR\alpha$ was found in most if not all cells in RC cell cultures from early proliferation stages through mineralized nodule formation. $ER\alpha$ was also detected in RC cells at all times analysed but at lower levels than $ERR\alpha$ which is also the case in rat bone marrow cultures
30 (data not shown). $ER\beta$, on the other hand, was more difficult to detect at any time other than in early proliferating cultures. These observations fit with the expression pattern of these three receptors in vivo in 21 day fetal calvaria.

Indeed, $ERR\alpha$ is more highly and widely expressed than either $ER\alpha$ or $ER\beta$, being highly expressed in sutural cells and all identifiable osteoblasts and osteocytes. $ER\alpha$ is not highly expressed in nascent, but is detectable in more mature, osteoblastic cells. $ER\beta$, on the other hand, is more highly expressed in sutural and nascent osteoblasts. These results suggest that $ERR\alpha$ and $ER\alpha$ and/or $ER\beta$ are co-expressed in at least some osteoblastic cells, and that these receptors may act alone or together to regulate the expression of target genes in bone. It is also notable that $ERR\alpha$ protein is localized primarily in either the nucleus or the cytoplasm or both depending on the developmental stage of the osteoblast. This suggests that $ERR\alpha$ target genes and function may vary depending on the maturational stage of the osteoblastic cells.

In addition to its expression in fetal calvaria, $ERR\alpha$ is also highly expressed in adult calvaria and other fetal and adult bones, including long bones such as the femur. It is also throughout osteogenesis in adult rat bone marrow stromal cell cultures, suggesting that it may function throughout the lifetime of the organism and in all bones of the body irrespective of the developmental process by which they form, i.e., through an intramembranous or endochondral route. In adult quiescent bone, labeling appears highest in osteocytes, which are thought to be mechanosensors that send strain-related signals to lining cells located at the bone surface through the canicular syncytium (Huskes et al, 2000), leading to recruitment of osteoblasts amongst other effects. These data suggest that $ERR\alpha$ may function not only during fetal bone development but also in adult life in both bone formation and maintenance.

Previously, it has been shown that the estrogen receptors $ER\alpha$ and $ER\beta$ are themselves expressed in osteoblasts and osteocytes (Braidman et al, 1995; Onoe et al, 1997; Windahl et al, 2000), raising the possibility that $ERR\alpha$ and one or both of the ERs may be co-expressed in at least some osteoblastic cells. The inventors have shown by both mRNA and protein analysis that $ERR\alpha$ and $ER\alpha$ are co-distributed in large cohorts of osteoblastic

cells, raising the possibility that these receptors may regulate the expression of the same target genes in bone via their known ability to participate in protein-protein interactions (Johnston et al, 1997) and their recently described capacity to bind to the same DNA target (SFRE and ERE) sequence on the osteopontin promoter (Vanacker et al, 1999).

Together, these data suggest that $ERR\alpha$, $ER\alpha$ and perhaps $ER\beta$ are co-expressed in osteoblastic cells, and may display at least some functions in common, either singly or through their interactions, with regulatory capacities to act on target genes.

$ERR\alpha$ and proliferation

Consistent with its expression in proliferating osteoblastic populations, we have found that antisense oligonucleotide-induced downregulation of $ERR\alpha$ inhibits proliferation of RC cell populations, an inhibition that appears to have consequences on bone nodule formation at later times (see below). The decrease in proliferation was somewhat unexpected, given our previous observation that $ERR\alpha$ expression appears to correlate with exit from proliferation and the onset of the differentiation process in at least certain other cell types, including the nervous system, the epidermis and muscles in the developing mouse (Bonnelye et al, 1997). This suggests that $ERR\alpha$ may play cell-type specific functions and is in keeping with its detection from the onset of osteogenesis in vivo (Bonnelye et al, 1997) and its presence in all osteoblastic cells including the earliest detectable osteoprogenitors (current data).

The molecular basis for the $ERR\alpha$ effect on proliferation is of interest. Since OPN has been described as a target gene of $ERR\alpha$ in *in vitro* promoter-reporter assays (Bonnelye et al, 1997; Vanacker et al, 1998), and since OPN is highly expressed in many proliferative populations including osteoprogenitors (Fig. 2) and in many tumour cell lines (see review in Denhardt and Noda, 1998), one candidate target in the proliferation time window for antisense-induced downregulation was OPN. However, we found

no detectable downregulation of this molecule during this developmental time window, although it is clearly sensitive to regulation by $ERR\alpha$ later during the differentiation phase of the cultures (see below). We also found no significant changes in the antisense-treated RC populations in expression of a variety of proliferation and apoptosis/survival-associated genes expressed in osteoblasts including c-fos, Bcl-2, and Bax. However, we did observe a significant decrease in cyclin D1, a regulator of G1 phase progression. Interestingly, estrogen induces cell proliferation by stimulating progression through the G1 phase of the cell cycle (Clarke et al, 1992; Wakeling et al, 1991), and induction of cyclin D1 expression is a critical feature of the mitogenic response to estrogen. There is also a strong correlation between increased levels of cyclin D1 mRNA with estrogen receptor overexpression in breast cancer cells (Buckley et al, 1993). Recently, Sabbah et al., have described a region in the cyclin D1 promoter that confers regulation by estrogens in the human mammary carcinoma cells MCF7. The induction is strictly hormone dependant and requires the DNA binding domain as well as both AF-1 and AF-2 domains of $ER\alpha$ (Sabbah et al, 1999). Although no ERE has been identified in the cyclin D1 promoter, it is possible that $ER\alpha$ activates cyclin D1 transcription by its ability to heterodimerize with c-jun/ATF-2 (Sabbah et al, 1999). $ERR\alpha$ has also been described as a modulator of the estrogen receptor-mediated response of the human lactoferrin gene promoter (Yang et al., 1996), a mechanism that may also underlie its ability to regulate cyclin D1.

25 $ERR\alpha$, osteoblast differentiation and matrix mineralization

The findings described herein show a critical role for $ERR\alpha$ in bone formation, with both up- and down-regulation of bone nodule formation concomitant with up- and down-regulation of $ERR\alpha$ expression *in vitro*. Up-regulation of $ERR\alpha$ by transfection of RC cells with a full-length $ERR\alpha$ expression vector late in the proliferation time window increased bone nodule formation by an amount approximately equivalent to the transfection efficiency

of the population. Concomitantly, all bone markers expressed at early differentiation stages (ALP, OPN, BSP and COLLI; Aubin and Liu, 1996) were upregulated 72h after transfection; OCN, a late marker of the mature osteoblast, was also upregulated at days 10 and 13. Whether the increase in
5 osteoprogenitor differentiation and bone nodule formation is a consequence of upregulation of any of these bone markers, or results from regulation of another currently unknown $ERR\alpha$ target gene, remains to be explicitly tested.

Downregulation of $ERR\alpha$ also had marked inhibitory effects on differentiation and bone nodule formation, when it was downregulated during
10 proliferation phase or earlier or later in the differentiation sequence. When RC cells were treated with antisense oligonucleotides only during the proliferation window (day 1-6) and then returned to normal medium, the number of bone nodules present at day 15 was reduced compared to untreated or sense-treated cells. While one can speculate that this decrease reflects the
15 downregulation of cyclin D1 and decreased proliferation of osteoprogenitors amongst other cells, the decrease could also reflect the concomitant downregulation of the bone "master" gene *Cbfa1* (Ducy et al, 1997; Komori et al, 1997) and BSP, both of which are upregulated early during osteoprogenitor cell differentiation (Aubin and Liu, 1996; Malaval et al., 1999); we also found
20 that BSP and $ERR\alpha$ are co-expressed in these very early osteoprogenitors (Fig. 2). The latter possibility is consistent with the finding that downregulation of $ERR\alpha$ only after proliferation has largely ceased (antisense treatment from day 5-11) results in complete inhibition of mineralized bone nodule formation, and concomitant downregulation of *cbfa1*, BSP and OCN. These
25 observations, together with the data on increased bone formation when $ERR\alpha$ is upregulated early, suggest that at least part of the effect of $ERR\alpha$ on osteoblast differentiation and bone formation occurs early during the differentiation sequence, such that differentiation may not progress beyond a certain point when $ERR\alpha$ levels are low. In keeping with this hypothesis,
30 large flat but ALP-positive colonies are present in antisense-treated cultures and a few cells express diminished levels of other osteoblast markers (Fig. 8).

It is also notable that $ERR\alpha$ also plays a role late in the differentiation/maturation sequence, i.e., when matrix is mineralizing. When RC cells were treated with antisense during late differentiation-matrix mineralization stages (day 9-15), we observed a less pronounced but
5 nevertheless dose-dependent decrease in mineralized nodule number, but those that did form appeared to cover a larger surface area (i.e., more bone was deposited per colony) than those in control cultures.

**$ERR\alpha$ expression is stimulated by estrogen in vitro and in vivo and is
10 upregulated in the OVX rat model of postmenopausal osteoporosis**

Estrogen (17β -estradiol; E2) was found to regulate $ERR\alpha$ at early times in chronically treated RC cell cultures, while an acute (24h) treatment at either day 9 or day 15 did not. These results suggest a link between $ERR\alpha$ and E2 in bone, most likely during the proliferation phase. Shigeta et al. showed that
15 E2 can also activate $ERR\alpha$ in the uterus (Shigeta et al, 1997). Importantly, we found that 17β -estradiol also upregulates $ERR\alpha$ in bone *in vivo*. Based on the kinetics of upregulation after E2 administration, this regulation appears to be an immediate or acute response to the administration of E2. Together, these results suggest a link between $ERR\alpha$ and estrogens in two estrogen-
20 sensitive tissues.

$ERR\alpha$ and $ER\alpha$ are both expressed in adult osteocytes in calvaria and long bones suggesting a function of $ERR\alpha$ during adult life. Bone loss in the aging skeleton is accelerated by a decrease in secretion of estrogens in postmenopausal women and can be reversed by administration of natural or
25 synthetic estrogens. The decrease in estrogen also induces a decrease in the expression of $ER\alpha$ (Hoyland et al, 1999). Given these data, it was surprising to measure an increase in $ERR\alpha$ in OVX rats, although the acute nature of the response in vitro to estrogen supplementation, similar to the acute upregulation in vivo which was followed by downregulation, may help to
30 explain the discrepancy. However, it is also worth noting that $ERR\alpha$ is highly expressed in the osteoblasts present in the high turnover bone of the OVX

rats, which may indicate that $ERR\alpha$ expression is essential for osteoblast function in osteoporosis. The data are consistent with E2 having a biphasic effect on $ERR\alpha$ expression in bone.

5 $ERR\alpha$ expression is stimulated by $TGF\beta$ and vitamin D3, decreased by PTH-1-34 and unaffected by dexamethasone in proliferating RC cell cultures

$ERR\alpha$ is also regulated during the proliferation stages of RC cell cultures by another hormone ($1,25(OH)_2D_3$) and a growth factor ($TGF\beta$ with
10 potent regulatory activities in bone metabolism. Acute treatment with $1,25(OH)_2D_3$ or $TGF\beta$ for 24 hours later in the developmental sequence (at day 9 or day 15) had no effect on $ERR\alpha$ expression. It will therefore be of interest to determine whether $1,25(OH)_2D_3$ and $TGF\beta$ effects may be mediated at least in part by its abilities to modify $ERR\alpha$ levels. There was also a down
15 regulation of $ERR\alpha$ by PTH-1-34 during proliferation which was most evident late in RC cultures when mineralized nodules are present. However, acute treatment with PTH-1-34 for 24 hours at day 7 or day 16 did inhibit the expression of $ERR\alpha$ which suggests a more direct effect of PTH-1-34 on $ERR\alpha$ expression compared to E2, $1,25(OH)_2D_3$ or $TGF\beta$.

20 $ERR\alpha$ expression in RC and RBM cell populations is almost the same in cells grown without (-Dex) or with (+Dex) dexamethasone. A 24 hour acute treatment of RC cells with Dex at day 9 or day 15 also had no effect on the $ERR\alpha$ expression level. These results suggest that, in RC and RBM cell populations, Dex has no effect on the expression of $ERR\alpha$ while it does
25 downregulate $ER\alpha$ and even more potently downregulates $ER\alpha$, suggest differential regulation by glucocorticoids of $ERR\alpha$ versus the ERs.

EXAMPLES

The examples are described for the purposes of illustration and are not
30 intended to limit the scope of the invention.

Methods of chemistry, molecular biology, protein and peptide biochemistry and immunology referred to but not explicitly described in this disclosure and examples are reported in the scientific literature and are well known to those skilled in the art.

- 5 The following materials and methods were employed in the Examples which follow.

Cell Culture

- 10 Cells were enzymatically isolated from the calvaria of 21 d Wistar rat fetuses by sequential digestion with collagenase as described previously (Bellows, 1986). Cells obtained from the last four of the five digestion steps (populations II-V) were pooled and plated in T-75 flasks in α -MEM containing 15% heat-inactivated FBS (Flow Laboratories, McLean, VA) and antibiotics comprising 100 mg/ml penicillin G (Sigma Chemical Co., St. Louis, MO), 50
- 15 mg/ml gentamycin (Sigma), and 0.3 mg/ml fungizone (Flow Laboratories). After 24 h incubation, attached cells were washed with PBS to remove nonviable cells and other debris, and then collected by trypsinization using 0.01% trypsin in citrate saline. Aliquots were counted with a Coulter Counter (Coulter Electronics, Hialeah FL), and the remaining cells were resuspended
- 20 in the standard medium described above. The resuspended cells were plated into 100 mm tissue culture dishes at 10^5 cells/dish, into 35mm tissue culture dishes at 2×10^4 /dish and in 24 wells plates at 10^4 cells/well. After 24 h incubation, medium was changed and supplemented with 50 μ g/ml ascorbic acid, 10 mM sodium β -glycerophosphate, and with or without 10^{-8} M
- 25 dexamethasone (Merck, Sharp, and Dohme, Canada, Ltd., Kirkland, PQ), or 10^{-8} M 17 β -estradiol (E2; Sigma), or 10^{-9} M 1,25(OH) $_2$ D $_3$, or 10^{-9} M PGE $_2$, 10^{-10} M TGF β or 10^{-11} M 1-34 PTH (Sigma). Medium was changed every 2 days. All dishes were incubated at 37°C in a humidified atmosphere in a 95% air/5% CO $_2$ incubator.

Bone marrow stromal cells from the femurs of young male Wistar rats, 110-120 g body weight, were cultured essentially as described (Aubin et al, 1998). The rats were killed by cervical dislocation, the femurs were dissected under aseptic conditions and placed in medium (MEM) containing antibiotics (1 mg/ml penicillin G (Sigma Chemical Co., St. Louis, MO), 500 µg/ml gentamycin sulfate (Sigma) and 3 µg/ml fungizone (Flow Laboratories, McLean, VA) (designated 10X AB)). The adherent connective tissue and muscles were removed, the femurs were placed in fresh 10X AB, and their ends (epiphyses and metaphyses) were cut off with a scalpel. With a 22 gauge needle and syringe, 5 ml of MEM were flushed through each femur until the bone appeared blanched (about five to eight times). The resulting cell suspension was flushed through a syringe several times to produce a largely single cell suspension; cells recovered from two femurs were added to a T-75 tissue culture flask (Falcon) and incubated in a 37°C humidified 95% air/5% CO₂ incubator. Growth medium, consisting of MEM containing 10% fetal calf serum, antibiotics (100 µg/ml penicillin G, 50 µg/ml gentamycin and 0.3 µg/ml fungizone), 50 µg/ml ascorbic acid, and 10⁻⁸ M dexamethasone (Sigma), was changed every 2-3 days. After 7 days, cells in each T-75 cell culture flask were washed with 15 ml of warm PBS and adherent cells were recovered with a mixture of 3 ml of 0.2% trypsin (w/v in citrate saline) and 2 ml of collagenase. Recovered cells were passed through a syringe with a 22 gauge needle to insure a single cell suspension. Cells were then counted on a Coulter Counter (Coulter Electronics, Hialeah FL) and plated at densities between 5X10³ and 2X10⁴ cells/35 mm dish and 2X10⁵ cells/100 mm dish (Falcon). Cells were cultured in αMEM supplemented as above and changed every 2-3 days, for approx. 17 days until mature bone nodules were seen. To promote mineralization, 10 mmol/ml of β-glycerophosphate (Sigma) was added for at least 2 days of culture prior to fixation. Cultures were then stained and the colonies quantified as indicated in what follows.

Northern blots

Total RNA was extracted with guanidine from RC cells at different times of the culture corresponding to different stages of proliferation, differentiation and bone nodule formation (Current Protocols in Molecular Biology, vol. 1, 1996). Northern blots were prepared and hybridized with a 750bp fragment corresponding to the rat 3' UTR of $ERR\alpha$ according to standard procedures (Chirgwin et al, 1979). Rat $\alpha 1$ COLL-I (Genovese et al.) was a 900bp cDNA PstI fragment containing the entire 3' noncoding region and one-half of the C-terminal of the propeptide of the $\alpha 1$ chain of type I. Rat bone/liver/kidney ALP (Noda et al., 1987) was a 600 bp cDNA *EcoRI* fragment obtained by digesting pRAP54 with *BssHII-XhoI* to remove 1.8 kb of the 5' region and religating the blunt ends. Rat OPN was a 700 bp cDNA *BamHI-EcoRI* fragment obtained by digesting the full length cDNA with *PvuI* to remove 800 bp of 5' region and ligating the blunt ended fragment into *SmaI* cut pGEM-7Zf(+) vector (Promega, Madison, WI). Rat OCN was a partial cDNA containing 350 bp of the 3' UTR isolated with OCN-specific primers from a λ gt11 library prepared from ROS 17/2.8 cells, rat BSP was a partial cDNA containing 500bp of 3' region isolated with BSP-specific primers from a λ gt11 library prepared from RC cells forming bone nodules, and rat L32 was generated from RC cell mRNA by PCR using specific primers; the identities of the OCN, BSP and L32 probes were confirmed by sequencing (Liu et al., 1994).

RT-PCR

Samples of total cellular RNA (1.5-5 μ g) were reverse-transcribed using oligo dT and the first strand synthesis kit of SuperscriptTM II, Gibco BRL. PCR was performed with specific primers specific for $ERR\alpha$. Primers, located in different exons, were as follows:

$ERR\alpha$ upstream (3'UTR): CAG GAA AGT GAA TGC CCA GG

$ERR\alpha$ downstream (3'UTR): CTT TGC AGC AAA TAT ACA TT

$ER\alpha$ upstream (Dom D 5'): GAG CTG CCA ACC TTT GGC CAA GT

ER α downstream (Dom D 3') : TGA ACT TGA TCG TGG AGA TTC

ER β upstream (Dom D): AAA GCC AAG AGA AAC GGT GGG CAT

ER β downstream (Dom E): GCC AAT CAT GTG CAC CAG TTC CTT

L32 upstream: CAT GGC TGC CCT TCG GCC TC

5 L32 downstream: CAT TCT CTT CGC TGC GTA GCC

The PCR reaction mixture contained cDNA (1 μ l), 1 μ l dNTP mix (20mM), 10X PCR buffer, Q solution, 25pmol primers and 5Units of Taq polymerase from Quiagen. PCR was done for 25 cycles (94°C for 1 min, 55°C for 1min, 72°C for 1min and a final elongation step of 7 min at 72°C) for ER α and L32; 35
 10 cycles (94°C for 1 min, 55°C for 1min, 72°C for 1min and a final elongation step of 7 min at 72°C) for ER α ; 45cycles (94°C for 1 min, 59°C for 1min, 72°C for 1min and a final elongation step of 7 min at 72°C) for ER β . Amplimers were sequenced for verification.

Osteoblast-associated and other markers were also amplified by PCR
 15 using specific primers for rat OCN, OPN, ALP, BSP, Cbfa1, COLL I (collagen type I α chain), C-fos, Cyclin D1, Bax and Bcl-2. PCR was done for 25 cycles (94°C for 1 min, 55°C for 1min, 72°C for 1min and a final elongation step of 7 min at 72°C) for OCN, OPN, ALP, BSP, L32, Bax and 30 cycles for Bcl-2, 32 cycles for c-Fos, 35 cycles for Cyclin D1 and Cbfa1 (with annealing
 20 temperatures of 58°C and 62°C respectively and 23 cycles for COLLI with annealing temperature of 59°C.

OC upstream: AGG ACC CTC TCT CTG CTC AC

OC downstream: AAC GGT GGT GCC ATA GAT GC

BSP upstream: CGC CTA CTT TTA TCC TCC TCT G

25 BSP downstream: CTG ACC CTC GTA GCC TTC ATA G

ALP upstream: CCC GCA TCC TTA AGG GCC AG

ALP downstream: TAG GCG ATG TCC TTG CAG C

OPN upstream: GCC ACT TGG CTG AAG CCT G

OPN downstream: GAA ACT CCT GGA CTT TGA CC

30 Cbfa1 upstream: CTT CAT TCG CCT CAC AAA C

Cbfa1 downstream: CAC GTC GCT CAT CTT GCC GG

Cyclin D1 upstream: TCC CGC CAG CAG CAA GAC AC
Cyclin D1 downstream: TGA GCT TGT TCA CCA GAA GC
c-Fos upstream: ATA GAG CCG GCG GAG CCG CG
c-Fos downstream: AAG CCC CGG TCG ACG GGG TG
5 Bax upstream: CCT TGG AGC AGC CGC CCC AG
Bax downstream: ATG TGG GCG TCC CGA AGT AGG
Bcl-2 upstream: GGG GAA ACA CCA GAA TCA AG
Bcl-2 downstream: AGA GAA GTC ATC CCC AGC CC
COLLI upstream: GGA GAG AGT GCC AAC TCC AG
10 COLLI downstream: CCA CCC CAG GGA TAA AAA CT

Poly (A) PCR Library selection

Nineteen poly(A)PCR libraries representative of five transitional stages in osteoblast lineage progression were selected from more than one hundred
15 available amplified colonies (Liu et al., 1994; Liu, F and Aubin, J.E., submitted). Stage A are replica-plated monolayer colonies committed to differentiate to the osteoblast lineage but not yet expressing type I $\alpha 1$ collagen or alkaline phosphatase, both early markers of osteoprogenitor cells. Stage B and C colonies are progressively more mature, i.e. expressing type I $\alpha 1$
20 collagen or both type I $\alpha 1$ collagen and alkaline phosphatase respectively. Stage D colonies represent multilayered cells and contain histologically identifiable cuboidal osteoblasts. Stage E colonies comprised terminal differentiation stages, with multilayered cells and mineralized bone matrix. Relative amounts of total cDNA were determined by Southern hybridization
25 and were used for normalization.

Western blots

Total protein was extracted from confluent HeLa and MC3T3-E1 cells according to standard methods (Current Protocols in Molecular Biology, vol. 1,
30 1996). Western blot analyses were performed using a semi-dry system. Immunoblotting was performed with rabbit polyclonal antiserum prepared against a rat peptide (NH-CPASDECEITKRR-C) localized in the C domain of

ERR α ; blots were incubated overnight at room temperature with the antiserum diluted to 1/500, and binding was detected using HRPO -conjugated goat-anti-rabbit antibodies (1/3000; BioRad) and chemoluminescence.

5 Immunolabelling

Immunolabelling of cultures was done essentially as described previously (Turksen, 1991; Turksen, 1992). Cultures were rinsed with PBS, fixed with 3.7% formaldehyde in PBS and permeabilized with methanol at -20°C. Frozen sections were fixed 10 min in cold acetone. Paraffine sections
10 were treated deparaffined in xylene, then rehydrated in 100%, 95% and 70% ethanol and water. After rinsing, cells in dishes or frozen, paraffine sections were incubated for 1 h at room temperature with 10% normal serum in PBS for ERR α and ER α and in 3% BSA in PBS (denaturated) for ER β OCN, ALP, OPN, and BSP. After rinsing, cells or sections were incubated for 1.5 hours
15 with appropriate dilutions of primary antibodies (1/50, anti-ERR α ; anti-ER α or anti- ER β (MC-20 or Y-19, respectively; Santa Cruz Biotechnology, Inc))(Shim et al, 1999; Saji et al, 2000; Tremblay et al 1999). The anti-rat OCN antiserum was kindly provided by Dominique Modrowski (INSERM U349, Hopital Lariboisiere, Paris, France) and used at 1/100 dilution. The anti-OPN
20 MPIIB10) and anti-BSP (WWVIDI) antibodies were purchased from the Hybridoma Bank (Iowa City, IA) and used at a 1/800 and 1/500 dilution respectively. The production and characterization of monoclonal antibody RBM 211.13 directed to rat bone/liver/kidney ALP, have been described elsewhere (Turksen and Aubin, 1991; Turksen et al., 1992); it was used at a
25 1/100 dilution of purified ascites fluid. 10% normal serum in PBS or 3% BSA in PBS were used as negative controls. Nodules or calvaria sections were rinsed in PBS and incubated for 1h at room temperature with secondary antibody CY-3-conjugated anti-rabbit (Jackson ImmunoResearch Lab, West Grove, PA, USA; 1/300 final dilution) for ERR α , OC and ALP or secondary
30 antibody anti-mouse (Jackson ImmunoResearch Lab, West Grove, PA, USA; 1/300 final dilution) for BSP and OPN. After rinsing, samples were mounted

in Moviol (Hoechst Ltd, Montreal, PQ, Canada) and observed by epifluorescence microscopy on a Zeiss Photomicroscope III (Zeiss, Oberkochen, Germany). For photography and printing, equal exposure times were used for specifically-labelled and control cultures.

5

Nodule quantification

For quantification of nodule formation, dishes or wells were fixed and stained by the Von Kossa technique and bone nodules were counted on a grid (Bellows et al., 1986; Bellows and Aubin, 1989). Results are plotted as the mean number of nodules \pm SD of three wells for controls and each concentration of antisense or sense primers and five dishes for pcDNA3 control and pcDNA3-ERR α .

10

Cell counting

For cell growth analysis, the cell layers were rinsed in PBS, released with trypsin and collagenase (1:1, vol/vol, of solutions described above), and the harvested cells were counted electronically. Results are plotted as the average of three counts for each of three dishes for control and pcDNA3-ERR α or three wells for each concentration of antisense or sense primers used.

20

Alkaline phosphatase histochemistry

The histochemical stain for alkaline phosphatase is a modification of Pearse's (1960). Cells were rinsed once with cold PBS and fixed in 10% cold neutral buffered formalin for 15 min, rinsed with distilled water, and left in distilled water for 15 min. Fresh substrate (10 mg Naphthol AS MX-PO4 (Sigma) dissolved in 400 μ l N,N-dimethylformamide, then added to 50 ml distilled water and 50 ml Tris-HCl (0.2 M, pH 8.3) and then 60 mg Red Violet LB salt (Sigma)), was filtered through Whatman's No. 1 filter directly onto the dishes, and incubated for 45 min at 20°C. The dishes were then rinsed in tap water, drained and stained with 2.5% silver nitrate for 30 min at room

25

30

temperature (von Kossa stain), then rinsed 3 times with tap water. The dishes were finally air dried.

Transient transfections

5 Primary RC cells were grown in 35mm tissue culture dishes at 2×10^4 /dish in α -MEM containing 10% heat-inactivated FBS (Flow Laboratories, McLean, VA) and supplemented with 50 μ g/ml ascorbic acid, 10 mM sodium β -glycerophosphate, and 10^{-8} M dexamethasone. Cells were transfected at 50% of confluence according to the Effecten transfection protocol (Quiagen) using a pcDNA3 empty plasmid as a control and pcDNA3-ERR α (in the EcoRI
10 cloning site) at 0.5 μ g of total DNA per transfection. As control of transfection efficiency, we used a CMV- β Gal vector. Nodules were counted at day 15. mRNA was extracted at 72h, day 10 (beginning of nodules formation) and day 15 (mineralized nodules), after transfection.

15

Antisense and sense oligonucleotide treatment

 The resuspended RC cells were plated in 24 wells plates at 10^4 cells/well. Antisense oligonucleotide inhibition of ERR α expression was accomplished with a 20-base phosphorothioate-modified oligonucleotide, localized to the A/B domain. The ERR α antisense oligonucleotide sequence
20 was: 5'-TCACCGGGGGTTCAGTCTCA-3'. Control dishes were treated with the complementary sense oligonucleotide or no oligonucleotide. Preliminary experiments were done to determine effective oligonucleotide concentrations that were not toxic. 0.1 μ M to 5 μ M oligonucleotides were added directly to
25 cells either during the proliferation phase (days 1 to 6) and 0.5 μ M to 2 μ M oligonucleotides were added during the differentiation phase (day 5 (end of proliferation) to 11) or (day 9 (nascent nodule formation) to 15) in standard medium as above supplemented with 50 μ g/ml ascorbic acid, 10 mM sodium β -glycerophosphate, and 10^{-8} M dexamethasone. Medium was changed
30 every 2 days and fresh oligonucleotides were added. mRNA was collected at

day 6 for the proliferation experiments and at day 15 for the differentiation experiments. Nodules were counted at 15 days.

Ovariectomized rats and estrogen-treated mice

5 8 week-old female CBA-1 mice from the University of Bristol breeding colony were treated once weekly by subcutaneous injection with either vehicle (0.2ml corn oil; Sigma Chemical, Poole, Dorset, U.K.) or 500µg of 17β-estradiol (Sigma) as described (Samuels et al, 1999). This dose was chosen because it has previously been reported to cause maximal stimulation of
10 endosteal bone formation in female mice (Brain et al, 1993). Throughout the experiment, animals received a standard diet (rat and mouse standard diet, B&K Ltd., Humbside, U.K.) and water ad libitum, and were kept with a cycle of 12h light and 12h darkness. Animals were sacrificed 4, 8, 12 and 24 days after the first subcutaneous injection, the tibia were removed for processing,
15 and mRNA was extracted (Samuel and Tobias, 1999).

 35-40 day old (100-125 g) Wistar rats (Charles River Breeding Laboratories, Quebec), either ovariectomized (OVX) or sham-operated (SHAM), were kept under standard diet and laboratory conditions for 4 weeks post-operation. Animals were killed, the uteri were dissected and weighed,
20 and the femurs were removed, fixed in 4% paraformaldehyde, decalcified for 2 weeks in EDTA and processed overnight for paraffin embedding.

Example 1

**ERRα mRNA and protein are expressed throughout all osteoblast
25 proliferation and differentiation stages *in vitro***

 ERRα mRNA expression levels were assessed over a proliferation-differentiation time course by Northern blotting of primary rat calvaria (RC) cell populations grown in the presence (+Dex) or absence (-Dex) of dexamethasone (Dex), a stimulator of differentiation in this model. Under
30 both growth conditions, ERRα mRNA was expressed at all times assessed, including proliferation (day 6), early nodule formation (day 10) and nodule mineralization (day 15) (Fig. 1A, B). For comparison, mRNA levels for three

osteoblast markers, alkaline phosphatase (ALP; a relatively early marker of osteoblast development), osteopontin (OPN) and osteocalcin (OCN; a late marker of osteoblast maturation), are also shown (Fig.1 A, B).

Because RC cell cultures comprise a heterogeneous mixture of cell types and osteoblasts at different differentiation stages, we sought next to confirm that $ERR\alpha$ is expressed by osteoblast lineage cells. To do this, we used globally-amplified (poly(A) PCR) cDNA pools prepared previously from single isolated osteoblast colonies at different stages of differentiation (Liu et al., 1994; Liu and Aubin, submitted). Colonies used were selected based on their molecular phenotypes (relative expression levels of collagen type I (COLL I), OPN, bone sialoprotein (BSP), ALP and OCN). $ERR\alpha$ was amplified in each cDNA pool with specific primers for sequences in the 3' UTR of $ERR\alpha$ and found to be expressed at all developmental times. Notably, however, levels were generally lower in more primitive progenitors (A) and osteoblastic cells associated with mineralized nodules (E), and higher in more mature precursors (B), preosteoblasts (C) and osteoblasts (D) (Fig. 2).

Immunocytochemistry was performed to determine whether $ERR\alpha$ protein is expressed in RC cell cultures. A Western blot of HeLa cell extracts was used to confirm the specificity of the $ERR\alpha$ antiserum. A single immunoreactive band was detected at 53Kd (Fig. 3A). In extracts of the control osteoblastic cell line, MC3T3-E1, $ERR\alpha$ was almost undetectable, however, a strong and single band at 53kd was observed in cell extracts of the same line transfected with an $ERR\alpha$ expression vector (Fig. 3B).

$ERR\alpha$ protein was found widely distributed in most, if not all, cells in RC cell cultures at all times analysed, including early proliferation stages (Fig. 3C), confluence (Fig. 3D), when nascent nodules were forming (Fig. 3E) and when nodules were mineralizing (Fig. 3F, G). Note especially, however, that staining for $ERR\alpha$ was more intense in the osteoblasts associated with both early and late bone nodules than in the surrounding fibroblastic cells (Fig. 3E, F). Interestingly, while $ERR\alpha$ is primarily found in the cytoplasm and perinuclear location at days 2 (Fig. 3C) and 4 (data not shown), by day 6 and

thereafter, including in mature osteoblasts, nuclear label is prominent (Fig. 3 E-G).

For comparison, protein expression of four osteoblast markers, ALP (Fig. 3I), BSP (Fig. 3J), OCN (Fig. 3K) and OPN (Fig. 3L), is also shown. As predicted, $ERR\alpha$ is co-expressed in osteoblasts with OPN, which was described earlier as a target gene of $ERR\alpha$ by cotransfection studies (Bonnelye et al 1997; Vanacker et al, 1998), supporting the hypothesis that $ERR\alpha$ may regulate OPN in osteoblasts *in vivo* as well. Clearly, $ERR\alpha$ is also highly co-expressed in cells with ALP, OCN and BSP.

Example 2

$ERR\alpha$ is also expressed in osteoblastic cells in vivo in fetal and postnatal rat calvaria

To extend the observations made in vitro to bones in vivo, immunocytochemistry was performed on sections of 21d. fetal rat calvaria, the same bones used for preparation of cell cultures. Consistent with the in vitro results, $ERR\alpha$ was found in all detectable cohorts of osteoblasts from those associated with nascent bone at the osteogenic front (Fig. 4 A) to those in the more mature growing bone trabecula (Fig. 4B) and remodeling bone (Fig. 4C). Consistent with the RT-PCR results on single bone nodules (cf. Fig. 2), $ERR\alpha$ was also detectable in sutural cells (arrows, Fig. 4A), preosteoblasts, osteoblasts (Fig. 4B, C) and osteocytes (Fig. 4B). $ERR\alpha$ was also high in the osteocytes present in postnatal (4 week) rat calvaria, suggesting that $ERR\alpha$ may be involved not only in the formation but also in the maintenance of bone. Also consistent with the in vitro cell labeling, $ERR\alpha$ in fetal calvaria in vivo is co-expressed in cells with ALP, OPN (Fig. 4 F, H), BSP (Fig. 4G) and OCN (Fig. 4I).

Example 3

Inhibition of $ERR\alpha$ expression blocks the proliferation of RC cells and their differentiation to mature bone-forming osteoblasts

Antisense oligonucleotides form DNA:RNA duplexes with specific mRNA species, thereby blocking binding of the mRNA to the 40S ribosomal subunit and preventing translation (Reddy et al., 1994). Preliminary experiments were done to determine effective oligonucleotide concentrations that were not toxic (not shown) and the specificity of the antisense was also confirmed by immunocytochemistry on bone nodules. After 24h of treatment or not with sense or antisense oligonucleotides, $ERR\alpha$ was detectable in bone nodules in untreated cultures and those treated with $1\mu M$ sense oligonucleotides but was almost undetectable in bone nodules present in cultures treated with $1\mu M$ antisense (data not shown).

To dissect the possible involvement of $ERR\alpha$ in osteoblast differentiation and bone formation, RC cells were treated at different developmental times from early proliferation stages until mineralized nodule formation.

When RC cells were treated between days 1-6, the proliferation stage, a significant and specific dose-dependent decrease in cell number (30% at $1\mu M$ and 40% at $2\mu M$), was found at day 6 in dishes treated with antisense compared to sense or untreated controls (Fig. 5A). These results suggest that $ERR\alpha$ may play a role in the proliferation or very early differentiation phases of RC cells. To analyze the underlying mechanism of $ERR\alpha$ action during the proliferation phase, expression of early markers of osteoblast differentiation (ALP, BSP, OPN, cbfa1, COLLI), proliferation (Cyclin D1, c-Fos) and apoptosis (Bcl2, Bax) were assessed at day 6 (Fig. 5B). BSP and cbfa1 were reduced significantly (Fig. 5C, D); Cyclin D1 was also reduced but it did not reach statistical significance. On the other hand, ALP, OPN, COLLI, c-Fos, Bax and Bcl2 were not significantly affected (Fig. 5C, D and data not shown). To determine if treatment during the proliferation time window caused a sustained alteration in differentiation, terminal differentiation/bone nodule formation was assessed at day 15 in cultures treated between days 1-6 with antisense and then switched to normal differentiation medium. A significant decrease in mineralized bone nodule number, i.e., 29% at $1\mu M$ and 45% at

2 μ M antisense oligonucleotides (Fig. 6A), was seen. Concomitantly, ALP and BSP expression remained lower than levels seen in control or sense-treated cultures, while OPN, OCN and COLLI were not significantly altered (Fig. 6B, C).

5 To determine whether $ERR\alpha$ also plays a role in osteoblast differentiation independently of an effect on proliferation, RC cells were treated with the antisense oligonucleotide beginning at day 5 (after cells had reached confluence and proliferation was decreased) to day 11. Although cell number was decreased slightly by day 15 (19% at 1 μ M and 35% at 2 μ M) in
10 antisense-treated cultures, a striking dose-dependent decrease in mineralized bone nodule formation was seen, i.e., 30% at 0.5 μ M, 60% at 1 μ M and 100% decrease at 2 μ M; the sense oligonucleotides had a small non-specific non-dose-dependent effect on nodule numbers. A similar inhibition of bone nodule formation was also observed when the osteoblastic cell line MC3T3-E1 was
15 treated with the antisense oligonucleotides over the comparable time period. In antisense-treated cultures, ALP-positive colonies were present and large in diameter but flat, suggesting that inhibition of $ERR\alpha$ blocked differentiation at an early stage such that progression to matrix deposition and maturation was reduced. Consistent with this interpretation, Cbfa1, BSP and OCN were all
20 decreased in antisense-treated cultures whereas OPN, COLLI and ALP were not affected. Immunocytochemistry confirmed the decrease in OCN- and BSP-expressing cells, but the maintenance of ALP expression in incipient bone nodules (data not shown).

To determine whether $ERR\alpha$ also plays a role at later stages, RC cells
25 were treated between days 9-15, when differentiation is well-progressed and nascent nodules are becoming three-dimensional. At this stage also, while sense oligonucleotides had a small non-specific, non-dose dependent effect on nodule number, antisense oligonucleotides caused a small but nevertheless dose-dependent and significant decrease in the number of
30 mineralized bone nodules formed, i.e., 17% at 1 μ M and 27% at 2 μ M. In parallel, OCN and OPN but not BSP, COLLI or ALP were decreased.

Interestingly, those bone nodules that did form in antisense-treated cultures appeared to cover a greater surface area compared to those in control cultures (data not shown).

5 **Example 4**

Overexpression of $ERR\alpha$ increases differentiation and bone nodule formation in RC cells

$ERR\alpha$ expression was upregulated by transient transfection of RC cells at day 5 when cells were between 50-60% confluent. By using a CMV- β Gal control vector, it was estimated that a maximal efficiency of transfection of 10-15% was obtained, which resulted in a 30% increase in $ERR\alpha$ levels observed on a Northern blot (Fig. 7A). At day 15, a significant increase (15%) in number of mineralized bone nodules was observed (Fig. 7B). In parallel, we assessed expression levels of osteoblast markers (ALP, OPN, BSP, COLLI, OCN) 72h after transfection, at day 10 when bone nodules had started to form and at day 15 when nodules are formed and mineralized. An increase in OPN was observed at 72 h and d 10 in cultures overexpressing $ERR\alpha$, consistent with previous data describing OPN as a target gene of $ERR\alpha$ in reporter assays (Bonnelye et al, 1997a, Vanacker et al, 1998) (Fig. 7C). COLL I was also increased at the early time points, suggesting a role for $ERR\alpha$ in the formation of a major component of the bone extracellular matrix. OCN, a mature osteoblast marker expressed only at later differentiation times, was undetectable at 72 h, but was increased at both day 10 and 15, whereas BSP was increased 72 h after transfection but not detectably altered at later times. Finally, ALP, which was increased 72h after transfection, was lower at days 10 and 15 (Fig. 7C).

Example 5

$ERR\alpha$ mRNA is more highly expressed in RC cell cultures than either $ER\alpha$ and $ER\beta$ mRNA and expression patterns vary

The expression of $ERR\alpha$ mRNA in RC cultures and single bone nodules prompted a comparison of the levels of its expression with those of the two estrogen receptors, $ER\alpha$ and $ER\beta$. When RT-PCR was done with primers specific for each of these three receptors, $ERR\alpha$ was found to be expressed at significantly higher levels than either $ER\alpha$ and $ER\beta$ and the two estrogen receptors were themselves present at different levels (i.e., $ERR\alpha$ was easily detected at 25 cycles, while 35 cycles and 40 cycles were required to detect $ER\alpha$ and $ER\beta$ respectively). In addition, the expression patterns of the three receptors over the proliferation-differentiation time course in RC cell cultures was strikingly different. Similarly to $ERR\alpha$, which decreased slightly, $ER\beta$ decreased markedly over time in -Dex cultures, whereas $ER\alpha$ increased (Fig. 8A, B). On the other hand, both $ERR\alpha$ and $ER\beta$ decreased over time in +Dex cultures, but $ER\alpha$ did not (Fig. 8A, B).

15 Example 6

$ERR\alpha$, $ER\alpha$ and $ER\beta$ proteins are also expressed in RC cultures, but only $ERR\alpha$ and $ER\alpha$ are detectable in bone nodules in vitro

To compare the in vitro localization of $ERR\alpha$ with the ER's in RC cultures, immunocytochemistry was performed with polyclonal antibodies specific for $ER\alpha$ and $ER\beta$ (Santa Cruz, CA; Shim et al, 1999; Saji et al, 2000; Tremblay et al, 1999) and for $ERR\alpha$.

Interestingly, $ER\alpha$ was detected in RC cells at all times analysed from early proliferation stages through nodule formation and mineralization with especially strong labeling of osteoblastic cells in nodules (data not shown). $ER\beta$, on the other hand, was more difficult to detect at any time other than in early proliferating cultures; in particular, $ER\beta$ was seldom detected in osteoblastic cells in bone nodules (data not shown).

ERR protein was found more widely distributed in RC cell cultures than either $ER\alpha$ or $ER\beta$. $ERR\alpha$ was found in most if not all cells in RC cell cultures from early proliferation stages through mineralized nodule formation. Interestingly, $ERR\alpha$ was found localized in the nucleus as was $ER\alpha$ in mature

osteoblasts. $ER\alpha$ was mainly nuclear from day 2 to day 6 but thereafter was cytoplasmic and nuclear, while $ER\beta$ was primarily perinuclear in all cells in which it could be detected (data not shown).

5 **Example 7**

$ERR\alpha$ is more highly and widely expressed *in vivo* in fetal rat calvaria than $ER\alpha$ and $ER\beta$

To extend the observations made *in vitro* to bones *in vivo*, immunocytochemistry was performed on 21d. fetal rat calvaria sections (data not shown). Consistent with the *in vitro* results, strikingly different expression patterns were seen for $ERR\alpha$, $ER\alpha$ and $ER\beta$. $ERR\alpha$ was found in all detectable cohorts of osteoblasts from those associated with nascent bone at the osteogenic front to those in more mature bone trabeculae including remodeling bone. $ER\alpha$, on the other hand, was not detected in any cells in the suture or osteogenic front, but was detected in some osteoblasts associated with more mature and remodeling bone. $ER\beta$ was detected in a pattern virtually reciprocal to that of $ER\alpha$, i.e., it was present in sutural cells and cells at the osteogenic front, but it was virtually undetectable in osteoblastic cells in more mature and remodeling bone, which is consistent with the mRNA expression in RC cells and the expression in bone nodules. Based on staining intensity, and in keeping with the RT-PCR results, $ERR\alpha$ was more highly expressed than either $ER\alpha$ and $ER\beta$ *in vivo*.

Example 8

25 **$ERR\alpha$ mRNA is also expressed throughout all osteoblast proliferation and differentiation stages in bone marrow stromal cell cultures *in vitro***

The calvaria bone, an example of a flat bone, forms by a process of intramembranous ossification, while many other bones form by a process of endochondral ossification. An example of a bone forming by endochondral ossification is the femur, and osteoprogenitor cells also reside in the bone marrow stroma of the femur. $ERR\alpha$ mRNA expression levels were therefore

also assessed over a proliferation-differentiation time course by Northern blotting of primary rat bone marrow (RBM) cell populations grown in the presence (+Dex) or absence (-Dex) of dexamethasone (Dex), a stimulator of differentiation in this model as it is in the calvaria-derived cells (data not shown). Under both growth conditions, $ERR\alpha$ mRNA was expressed at all times assessed, including proliferation (day 4), early nodule formation (day 6-9) and nodule mineralization (day 14-17) phases. There was a trend towards increased $ERR\alpha$ expression at early and very late stages, the latter being when mineralized nodules are present, but otherwise levels were relatively constant.

Example 9

$ERR\alpha$ mRNA is more highly expressed in RBM cell cultures than either $ER\alpha$ and $ER\beta$ mRNA and expression patterns vary

The differential expression of $ERR\alpha$, $ER\alpha$ and $ER\beta$ mRNA in RC cultures and single bone nodules prompted a comparison of the levels of the three receptors in RBM cultures as well. When RT-PCR was done with primers specific for each of the three receptors. As in the RC model, $ERR\alpha$ was expressed at significantly higher levels than either $ER\alpha$ and $ER\beta$ and the two estrogen receptors were themselves present at different levels (i.e., $ERR\alpha$ was easily detected at 25 cycles, while 35 cycles and 40 cycles were required to detect $ER\alpha$ and $ER\beta$ respectively) in the RBM model (Fig. 9A,B). In addition, the expression patterns of the three receptors over the proliferation-differentiation time course in RBM cell cultures was strikingly different. Whereas $ERR\alpha$ mRNA expression was relatively uniform over the time course analysed, with or without Dex in the medium, $ER\alpha$ increased late in the proliferation phase (day 4-9) and decreased thereafter, although levels remained higher than at earliest times in -Dex cultures. $ER\beta$ also increased during the proliferation phase, but peaked later than $ER\alpha$, i.e., at day 11 (early differentiation phase) and then also decreased. Dex appeared to have a small inhibitory effect on the peak levels reached by both $ER\alpha$ and $ER\beta$.

Example 10

ERR α is also expressed in osteoblastic cells in fetal and postnatal rat femur

5 To extend the observations made in vitro to bones in vivo, immunocytochemistry was performed on sections of 21d. fetal and adult rat femurs (data not shown). Consistent with the in vitro results, ERR α was highly expressed in osteoblasts associated with the growing trabecular and cortical bone. ERR α was also found in the osteocytes present in cortical bone, and in
10 the osteocytes present in the secondary ossification zone, trabecular bone and cortical bone in postnatal (24 days) rat femur; ERR α is also highly expressed in the bone marrow of these animals. These data suggest that ERR α may be involved not only in the formation but also in the maintenance of the bone of the axial skeleton.

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Example 11

ERR α expression is stimulated by estrogen, vitamin D3, and TGF β and inhibited by PTH-1-34 in proliferating RC cells cultures.

To determine whether other hormones or growth factors that influence
20 proliferation and differentiation in RC cell cultures may modulate ERR α levels, we treated RC cells continuously with estrogen (17 β -estradiol, E2; 10⁻⁸M), 1,25(OH)₂ vitamin D3 (10⁻⁹M), PGE₂ (10⁻⁹ M), TGF β (10⁻¹⁰ M) or PTH-1-34 (10⁻¹¹ M). E2 (40% increase), D3 (47% increase) and TGF β (400%) stimulated expression of ERR α mRNA at day 6, but not later (Fig. 10 A,B).
25 An acute exposure of RC cells to E2, D3 or TGF β for 24 hours beginning at day 9 (nascent nodule formation) or day 15 (mature nodules present) (E2 and D3) or day 7 and day 15 (TGF β had no effect on ERR α mRNA levels (Fig. 10, A, B). PTH-1-34 inhibited ERR α at day 6 (15%) and day 15 (26%), but increased (20%) it at day 12 (Fig. 10C). Interestingly, an acute exposure to
30 PTH-1-34 at day 7 or day 16 decreased ERR α by 25% and 33% respectively

(Fig. 10C). Neither PGE₂ nor Dex had any significant effect on ERR α expression.

Example 12

5 **ERR α expression is stimulated by estrogen in mouse femur in vivo**

ERR α expression was analysed in samples of mRNA extracted from total femurs or femurs from which bone marrow had been flushed of mice treated with 17 β -estradiol at doses known to elicit a large anabolic effect on endosteal bone (Samuels et al, 1999). In samples of total femoral mRNA, E2
10 had a small stimulatory effect on ERR α mRNA levels that was most evident days 2, 12 and 16 (Fig. 11B). In the mRNA extracted from femurs from which bone marrow had been removed, a marked stimulatory effect of E2 on ERR α levels was evident at day 1, 2 and 4 (Fig. 11A, B).

15 Example 13

ERR α expression is stimulated in ovariectomized rats

ERR α is expressed in adult osteocytes in calvaria and long bone, suggesting a function for ERR α during adult life and in diseases of bone, including e.g., those characterized by decreased bone mass such as
20 osteoporosis. Ovariectomized (OVX) rats are a model for human postmenopausal (estrogen loss-induced) osteoporosis. Four weeks after surgery, ERR α expression was increased in osteocytes in the secondary ossification zone and cortical bone (Fig. 12D,H) of long bone and osteoblasts associated with trabecular bone (Fig. 12H, arrows) in OVX compared to
25 sham-operated (Fig. 12A, C, E, G) rats. High expression of ERR α was also found in the abundant osteoclasts in OVX rats (Fig. 12I, arrows). As indicated above, ERR α is expressed in adult calvaria, a site thought not to be affected by OVX, and no striking differences were found in staining intensities in sections from sham versus OVX rats (Fig. 12J, K).

References

- Aubin, JE, and Liu, F. (1996) in Raisz LG, Rodan GA, Bilezikian, JP (eds)
5 Principles of Bone Biology. Academic Press, San Diego, pp1-45.
- Bain, SD, Bailey, MC, Celino, DL, Lantry, MM, Edwards, MW. (1993) J, Bone
and Mineral Research, 8:435-442
- 10 Barreback, E.D. (1995) Antibody Engineering, 2d. Edition, Oxford University
Press
- Bellow, C.G., and Aubin, J.E. (1989) Dev. Biol. 133, 8-13
- 15 Bellow, C.G., Aubin, J.E., Heersche, J.N.M. and Antosz, M.E. (1986) Calcified
Tissue Int., 38: 143-154.
- Bonnelye E, JM. Vanacker, X. Desbiens, T. Dittmer, A. Begue, J. Aubin,
V. Laudet and B. Fournier. Mol Endocrinol, 11, 1997: 905-916 (a)
- 20 Bonnelye E, JM. Vanacker, S. Alric, N. Spruyt, B. Fournier, X. Desbien and V.
Laudet .
MOD, 65, 1997: 71-85 (b)
- 25 Braidman, JP., Davenport, LK, Howard Carter, D, Selby, PL, Mawer. EB, and
Freemont, AJ, (1995) , LK. (1995) J, Bone and Mineral Research, 10:74-80
- Buckley, MF, Sweeny, KJE, Hamilton, JA, Sini, RL, Manning, DL, Nicholson, RI,
DeFazio, A Watts, CKW, Musgrove, EA and Sutherland, RL. (1993) Oncogene
30 8:2127-2133.
- Case, AM and Reid, RL (1998) Arch Intern Med 158 (13):1405-12.

Chirgwin, J.J., Przbyla, A.E., MacDonald, R.J., and Rutter, W.J. (1979) In Current CP protocols . pp.4.2.1-4,2.8 .John Wiley&Sons, Inc.

- 5 Clarke, R, Dickson, RB, and Lippman, ME. (1992) Crit. Rev. Oncol. Hematol. 12:1-23.

Da Silva, JA, Larbre, JP, Seed, MP, Cutolo, M, Villaggio, B, Scott, DL, Willoughby, DA (1994)

- 10 J Rheumatol 21 (2):330-7

Denhardt, D.T. and Guo, X. (1993) FASEB J., 7, 1475-1482

Denhardt, D.T and Noda, M (1998) J Cell Biochemistry Supp 30/31:92-102.

15

Ducy, P, Zhang, R, Geoffroy, V, Ridall, AL and Karsenty, G. (1997) cell 89 (5): 747-54.

Enmark, E and Gustafsson, J-A. (1996) Mol. Endocrinol., 10, 1293-1307

20

Eriksen, EF, Colvard, DS, Berg, NJ, Graham, ML, Mann, KG, Spelsberg, TC, and Riggs, BL 1988 Science 241:84-86

Genovese, C, Rowe, D and Kream, B (1984) Biochemistry 23: 6210-6216.

25

Giguère V, Yang N, Segui P, Evans RM 1988 Nature 331 : 91-94

Green, S., Walter, P., Kumar, V., Krust, A., Bornert, J.M., Argos, P. and Chambon, P. (1986) Nature, 320, 134-139.

30

Green,S. and Chambon,P (1991) Molecular Mechanisms, Cellular Functions, Clinical Abnormalities.Malcom G.Parker Ed., Academic Press, Harcourt Brace Jovanovich, Publishers, London.

- 5 Gronemeyer H, Laudet V 1995 Transcription factors 3: nuclear receptors. Protein profile. 2 : 1173-1308

Grynblas, MD, Huckell,CB, Reichs, KJ, Derousseau, CJ, Greenwood, C, Kessler MJ (1993) J, Bone and Mineral Research 8(8): 909-17

10

Harris,SA, Tau,KR, Turner,RT and Spelsberg,TC, (1996) in Raisz LG, Rodan GA, Bilezikian, JP (eds) Principles of Bone Biology. Academic Press, San Diego, pp507-520.

- 15 Hines,K.L, Christ, M and S.M Wahl. (1993) Immunomethods. 3:13-22.

Hong,H, Yang,L, and Stallcup,MR (1999) J. Biol. Chem, 274 (32): 22618-22626.

Horwitz et al., (1999), Nat. Med. 5(3): 309-313.

20

Hoyland,JA, Baris,C, Wood,L, Baird,P, Selby, PL, Freemont, AJ and Braidman, IP (1999)
J Patho 188 (3): 294-303

- 25 Huiskes,R, Ruimerman,R, Van Lenthe,GA and Janssen,JD. (2000) Nature 405: 704-706.

Johnston,S.D., X.Liu, F.Zuo, T.L.Eisenbraun, S.R.Wiley, R.J.Kraus and J.E.Mertz. Mol.Endocrinol. (1997) 11 : 342-352.

30

Korach,K.S. (1994). Science, 266, 1524-1527.

- Komori, T, Yagi,H, Nomura,S, Yamaguchi,A, Sasaki,K, Deguchi,K, Shimiz,Y, Bronson, RT, Gao,YH, Inada,M, Sato,M, Okamoto,R, Kitamura, Y, Yoshiki, S, and Kishimoto,T. (1997) cell 89(5):755-64.
- 5 Kuiper,G.G.JM,. Enmark,E., Pelto-Huikko,M., Nilsson,S. and Gustafsson,J.A. (1996) Proc.Natl Acad. Sci. USA, 93, 5925-5930.
- Komm, BS, Teruening, CM, Benz,DJ, Graeme, KA, Gallegos, A, Korc, M, Greene, GL, O'Malley BW and Haussler, MR 1988 Science 241:81-84.
- 10 Liu,F., Malaval, L., Gupta,A.K., and Aubin,J.E.(1994) Dev.Biol. 166,220-234.
- Liu,F.and Aubin,J.E. (2000) Submitted.
- 15 Malaval, L., Liu, F., Roche, P., and Aubin, J.E. (1999) J. Cell. Biochem., 74, 616-627.
- Masi,AT, Freigenbaum, SL and Chatterton,RT (1995) Semin Arthritis Rheum 25 (1):1-27
- 20 Noda,M., Yoon,K.,Thiede,M., Buenaga,R., Weiss,M., Henthorn,P., Harris,H., and Rodan,G. (1987) J.Bone Miner. Res.2,161-164.
- Onoe,Y, Miyaura,C, Ohta,H, Nozawa,S and Suda,T, (1997) Endocrinology 138 (10): 4509-4512.
- 25 Pacifici, R. (1996) J.Bone Miner.Res., 11, 1043-1051.
- Pettersson K, Svensson K, Mattsson R, Carlsson B, Ohlsson R, Berkenstam A 1996 Mech. Dev. 54 : 211-213
- 30

Reddy,SV, Takahashi,S Dallas M, Williams RE, Neckers L and Roodman GD.
(1994) J.Bone Miner.Res., 9, 753-757.

Rosner,IA, Goldberg, VM and Moskowitz, RM (1986) Clin Orthop 213:77-83

5

Sabbah,M, Courilleau, D, Mester,J and Redeuilh,G. (1999) Proc Natl Acad Sci
96:11217-11222.

Saji,S, Jensen,EV, Nilsson,S, Rylander,T, Warner,M and Gustafsson, JA. (2000)

10 Proc Natl Acad Sci

97 :337-342

Sambrook et al., (1989) Molecular Cloning: A Laboratory Manual, Cold Spring
Harbor Laboratory Press, Cold Spring Harbor, N.Y.

15

Samuels,A, Perry,M.J and Tobias,J.H (1999) J, Bone and Mineral Research, 14:178-
186.

Shigeta, H., Zuo, W., DiAugustine, R., and Teng C,T. (1997) J. of Molecular

20 Endocrinology. 19, 299-309.

Shim,WS, DiRenzo,J, DeCaprio, JA, Santen,RJ, Brown,M and Jeng,MH (1999) Proc
Natl Acad Sci 96:208-213

25 Sladek,R., J.A.Bader and V.Giguère. (1997) Mol.Cell.Biol. 17 : 5400-5409.

Stein et al. (2000) Cancer 88 (12 Suppl): 2899-2902.

Tremblay,A, Tremblay,GB, Labrie,F and Giguere,V (1999) Mol Cell 3:513-519.

30

Turksen, K., and Aubin,J.E. (1991) J.Cell Biol. 114, 373-384.

Turksen,K., Bhargava, U., Moe,H.K., and Aubin, J.E. (1992) J. Histochem. Cytochem.40,1339-1352.

Turner RT, Riggs BL, Spelsberg TC. (1994). Endocr. Rev. 15 : 275-300

5

Vanacker,J-M., Pettersson, K., Gustafsson, J-A., and Laudet, V. (1999). EMBO 18, 4270-4279.

Vanacker,J.-M., Delmarre,C., Guo,X. and Laudet,V. (1998) Cell Growth Differ., 9,1007-1014.

10

Vega, RB and Kelly, DP. (1997) J. Biol. Chem. 272 : 31693-31699.

Wakeling,A.E, Dukes,M and Bowler,J. (1991) Cancer Res. 51: 3867-3873.

15

Weiss,MJ, Cole,DE, Ray,K, Whyte, MP, Lafferty, MA, Mulivor,RA and Harris, H. (1988) Proc Natl Acad Sci 85(20):7666-9.

Wiley SR, Kraus RJ, Zuo F, Murray EE, Loritz K, Mertz JE (1993) Genes and Dev. 7: 2206-2219

20

Windahl,SH, Vidal,O, Andersson, G, Gustafsson, J-Aand Ohlsson (1999) J Clin Invest 104:895-901.

Windahl,SH, Norgard,M, Kuiper, GGJM, Gustafsson, J-A and Andersson, G, (2000) Bone 26: 117-121.

25

Xie,W, Hong,H, Yang, NN, Lin, RJ, Simon,CM, Stallcup,MR and Evans,RM. Mol Endocrinol (1999) 13(12);2151-62.

30

Yang N, Shigeta H, Shi H, Teng CT (1996) J. Biol. Chem. 271 : 5795-5804

Yang,C and Chen,S (1999) Cancer Res 15; 59 (18): 4519-4524.

Yang, C, Zhou, D and Chen, S (1998) Cancer Res 58 : 5695-5700.

Zhang,Z and Teng,T. (2000) J. Biol. Chem J 275: 20837-20846.

5

Zellar,R. and Rogers, M. (1989). In Current CP protocols . pp.14.3.1-14.3.14.
John Wiley&Sons, Inc.

THE EMBODIMENTS OF THE INVENTION IN WHICH AN EXCLUSIVE
PROPERTY OR PRIVILEGE IS CLAIMED ARE DEFINED AS FOLLOWS:

1. A method of increasing proliferation of osteoblasts in a mammal
5 comprising administering to the mammal an effective amount of an agent
selected from the group consisting of:
 - (a) an estrogen related receptor alpha (ERR α) agonist;
 - (b) a substantially purified ERR α protein; and
 - (c) a nucleotide sequence encoding ERR α protein.
 - 10 (d) an agent which enhances expression of a gene encoding an
ERR α protein.
2. A method of increasing differentiation of osteoblasts in a mammal
comprising administering to the mammal an effective amount of an agent
15 selected from the group consisting of:
 - (a) an ERR α agonist;
 - (b) a substantially purified ERR α protein; and
 - (c) a nucleotide sequence encoding ERR α protein.
 - (d) an agent which enhances expression of a gene encoding an
20 ERR α protein.
3. A method of reducing proliferation of osteoblasts in a mammal
comprising administering to the mammal an effective amount of an agent
selected from the group consisting of:
 - 25 (a) an ERR α antagonist;
 - (b) a purified antibody which binds specifically to an ERR α protein;
 - (c) an antisense nucleotide sequence complementary to and
capable of hybridizing to a nucleotide sequence encoding an
ERR α protein; and
 - 30 (d) an agent which reduces expression of a gene encoding an
ERR α protein.

4. A method of reducing differentiation of osteoblasts in a mammal comprising administering to the mammal an effective amount of an agent selected from the group consisting of:

- 5 (a) an $ERR\alpha$ antagonist;
- (b) a purified antibody which binds specifically to an $ERR\alpha$ protein;
- (c) an antisense nucleotide sequence complementary to and capable of hybridizing to a nucleotide sequence encoding an $ERR\alpha$ protein; and
- 10 (d) an agent which reduces expression of a gene encoding an $ERR\alpha$ protein.

5. A method for treating a disorder associated with bone loss in a mammal comprising administering to the mammal an effective amount of an agent selected from the group consisting of:

- 15 (a) an $ERR\alpha$ agonist;
- (b) a substantially purified $ERR\alpha$ protein; and
- (c) a nucleotide sequence encoding $ERR\alpha$ protein.
- (d) an agent which enhances expression of a gene encoding an
- 20 $ERR\alpha$ protein.

6. The method of claim 5 wherein the disorder is selected from the group consisting of osteoporosis, osteoarthritis, Paget's disease, periodontal disease, osteolytic bone tumour metastases in, for example, breast cancer and multiple
25 myeloma, osteochondrodysplasias, osteogenesis imperfecta, sclerosing bone dysplasias and osteomalacia.

7. A method for treating a disorder associated with unwanted bone formation comprising administering to the mammal an effective amount of an agent selected from the group consisting of:

- 30 (a) an $ERR\alpha$ antagonist;

- (b) a purified antibody which binds specifically to an $ERR\alpha$ protein;
 - (c) an antisense nucleotide sequence complementary to and capable of hybridizing to a nucleotide sequence encoding an $ERR\alpha$ protein; and
 - 5 (d) an agent which reduces expression of a gene encoding an $ERR\alpha$ protein.
8. The method of claim 7 wherein the disorder is selected from the group consisting of fibrodysplasia ossificans progressiva, osteoblastic bone
- 10 metastases such as prostate cancer and osteosarcomas.
9. The method of any of claims 1 to 8 wherein the mammal is a human.
10. A method for screening a candidate compound for its ability to
- 15 modulate $ERR\alpha$ activity comprising:
- (a) providing a system for measuring a biological activity of $ERR\alpha$; and
 - (b) measuring the biological activity of $ERR\alpha$ in the presence or absence of the candidate compound,
- 20 wherein a change in $ERR\alpha$ activity in the presence of the compound relative to $ERR\alpha$ activity in the absence of the compound indicates an ability to modulate $ERR\alpha$ activity.
11. The method of claim 10 wherein the system for measuring a biological
- 25 activity of $ERR\alpha$ is a rat calvaria cell culture or a bone marrow stromal cell culture.
12. A method for screening a candidate compound for potential efficacy in promoting bone formation comprising:
- 30 (a) providing an assay system for determining $ERR\alpha$ agonist activity of a compound; and

- (b) testing the candidate compound for $ERR\alpha$ agonist activity in the assay wherein $ERR\alpha$ agonist activity in the candidate compound indicates potential efficacy as a promoter of bone formation.

5 13. A method for screening a candidate compound for potential efficacy in inhibiting bone formation comprising:

- (a) providing an assay system for determining $ERR\alpha$ antagonist activity of a compound; and
- (b) testing the candidate compound for $ERR\alpha$ antagonist activity in
10 the assay

wherein $ERR\alpha$ antagonist activity in the candidate compound indicates potential efficacy as an inhibitor of bone formation.

15 14. A pharmaceutical composition comprising an effective amount of an agent selected from the group consisting of:

- (a) an $ERR\alpha$ agonist;
- (b) a substantially purified $ERR\alpha$ protein; and
- (c) a nucleotide sequence encoding $ERR\alpha$ protein
and a pharmaceutically acceptable carrier.
- 20 (d) an agent which enhances expression of a gene encoding an $ERR\alpha$ protein.

15 15. A pharmaceutical composition comprising an effective amount of an agent selected from the group consisting of:

- 25 (a) an $ERR\alpha$ antagonist;
- (b) a purified antibody which binds specifically to $ERR\alpha$ protein;
- (c) an antisense nucleotide sequence complementary to and capable of hybridizing to a nucleotide sequence encoding $ERR\alpha$ protein; and
- 30 (d) an agent which reduces expression of the gene encoding $ERR\alpha$ protein and a pharmaceutically acceptable carrier.

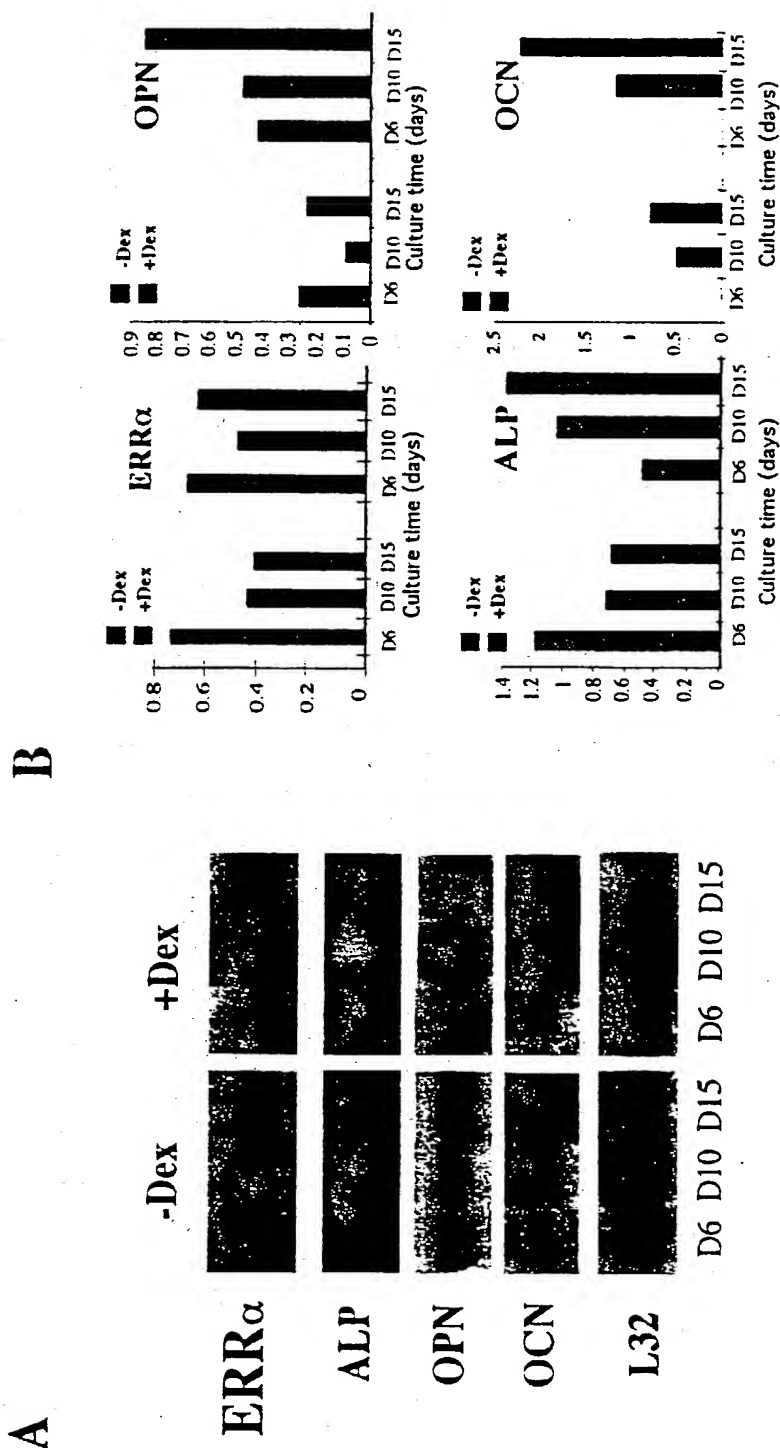


Figure1

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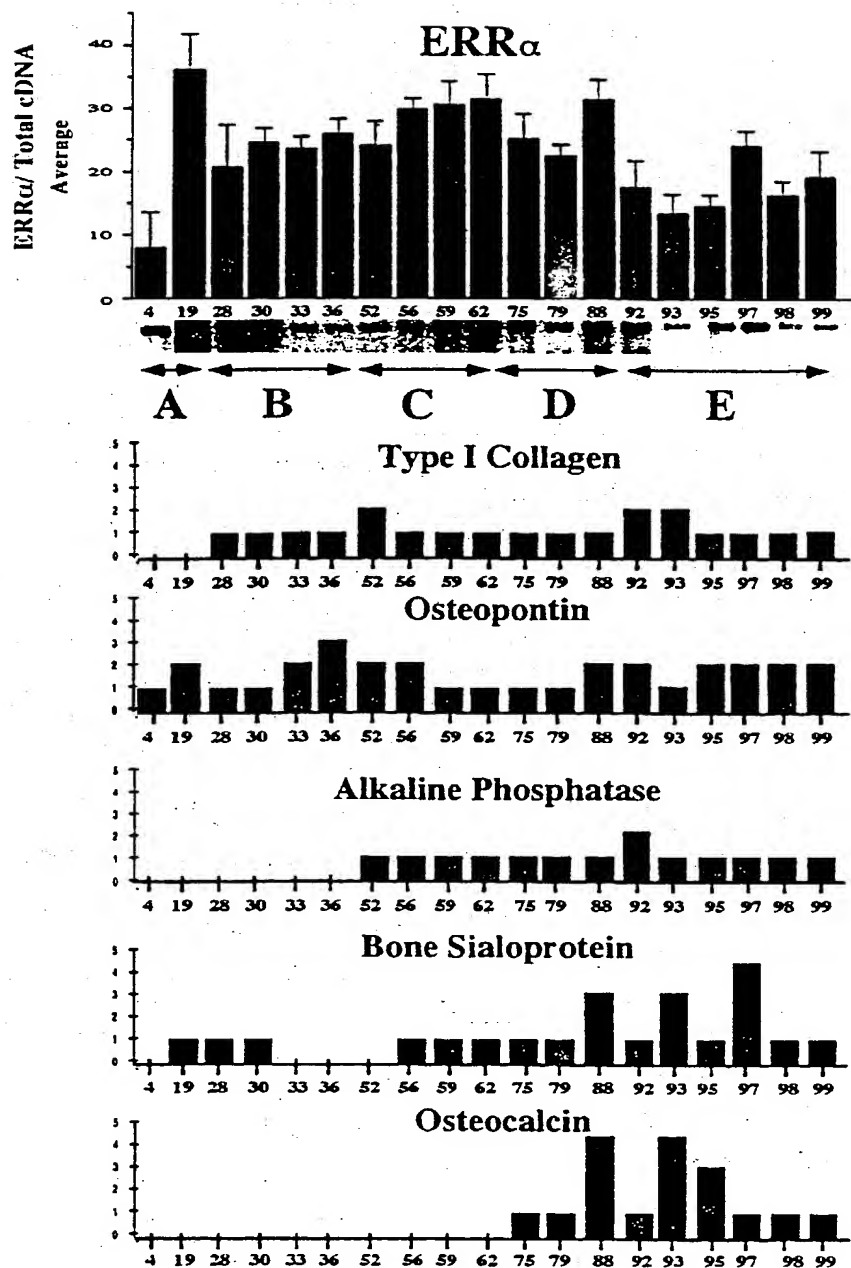


Figure2

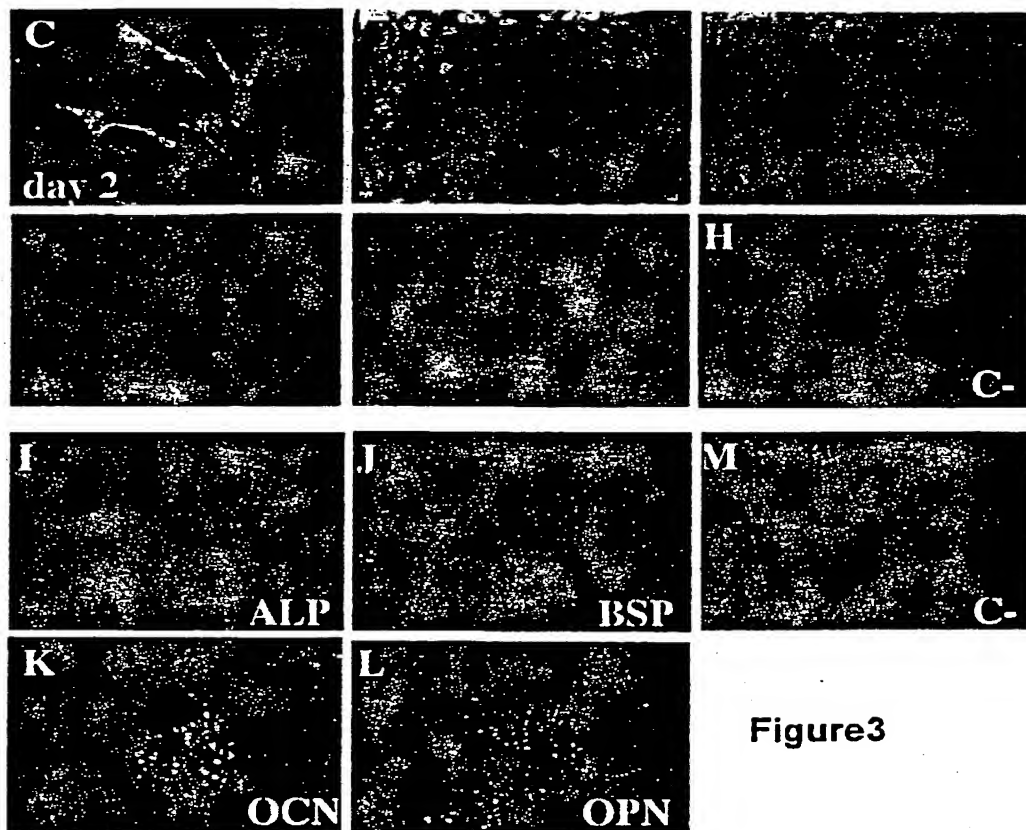
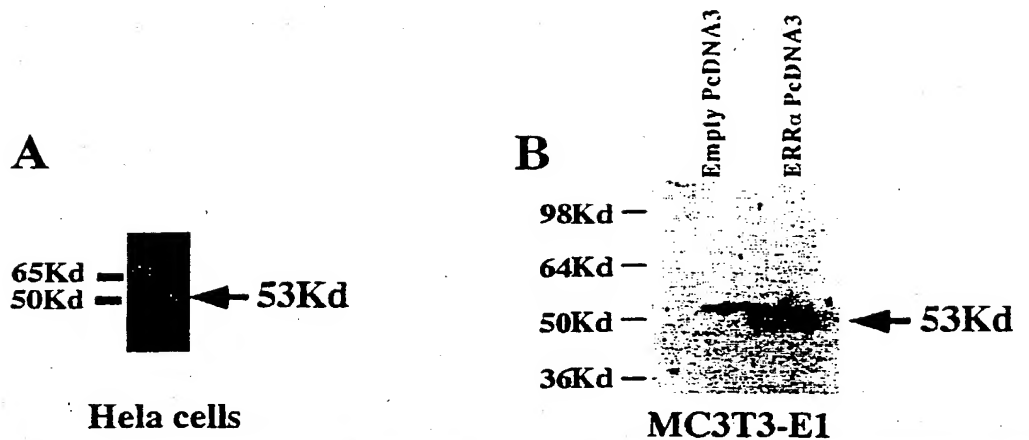


Figure3

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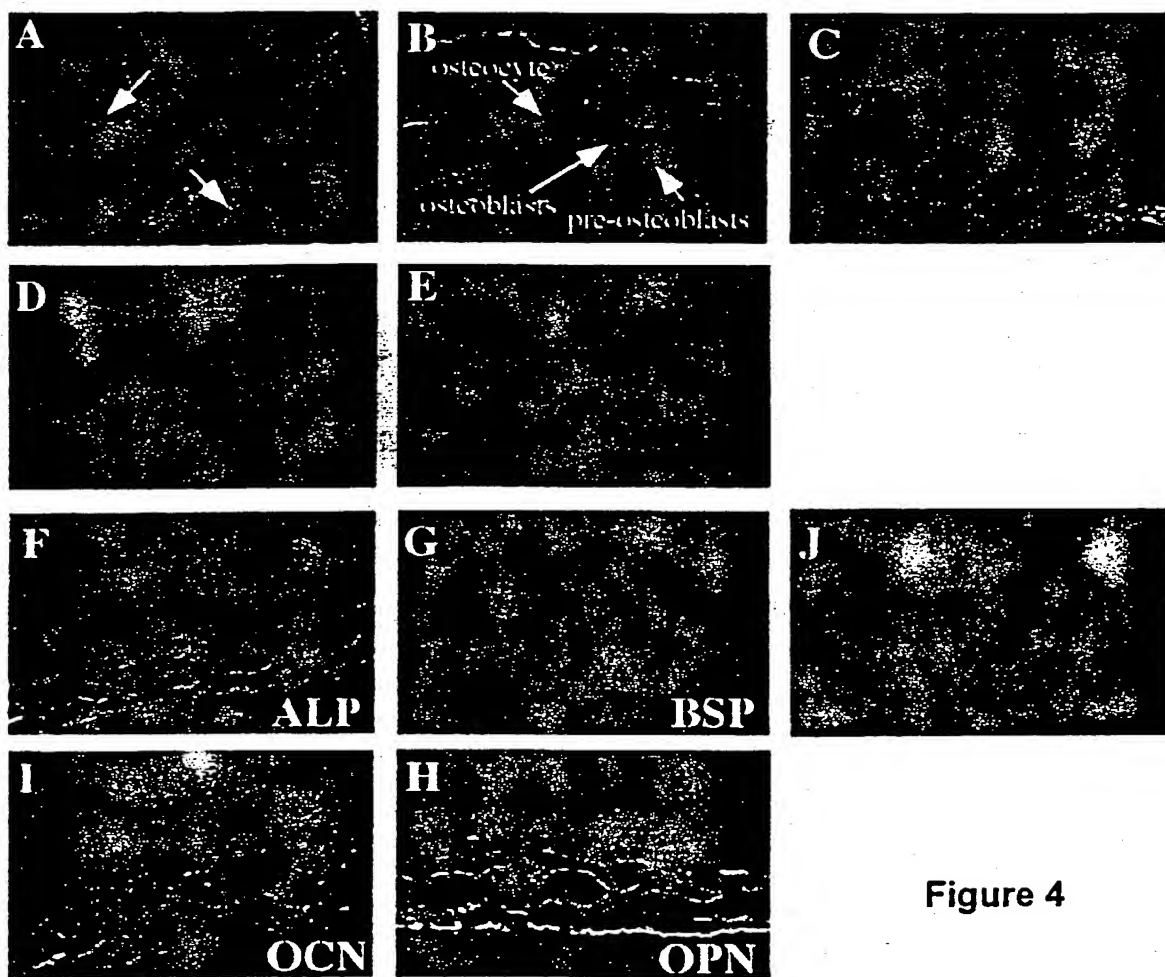
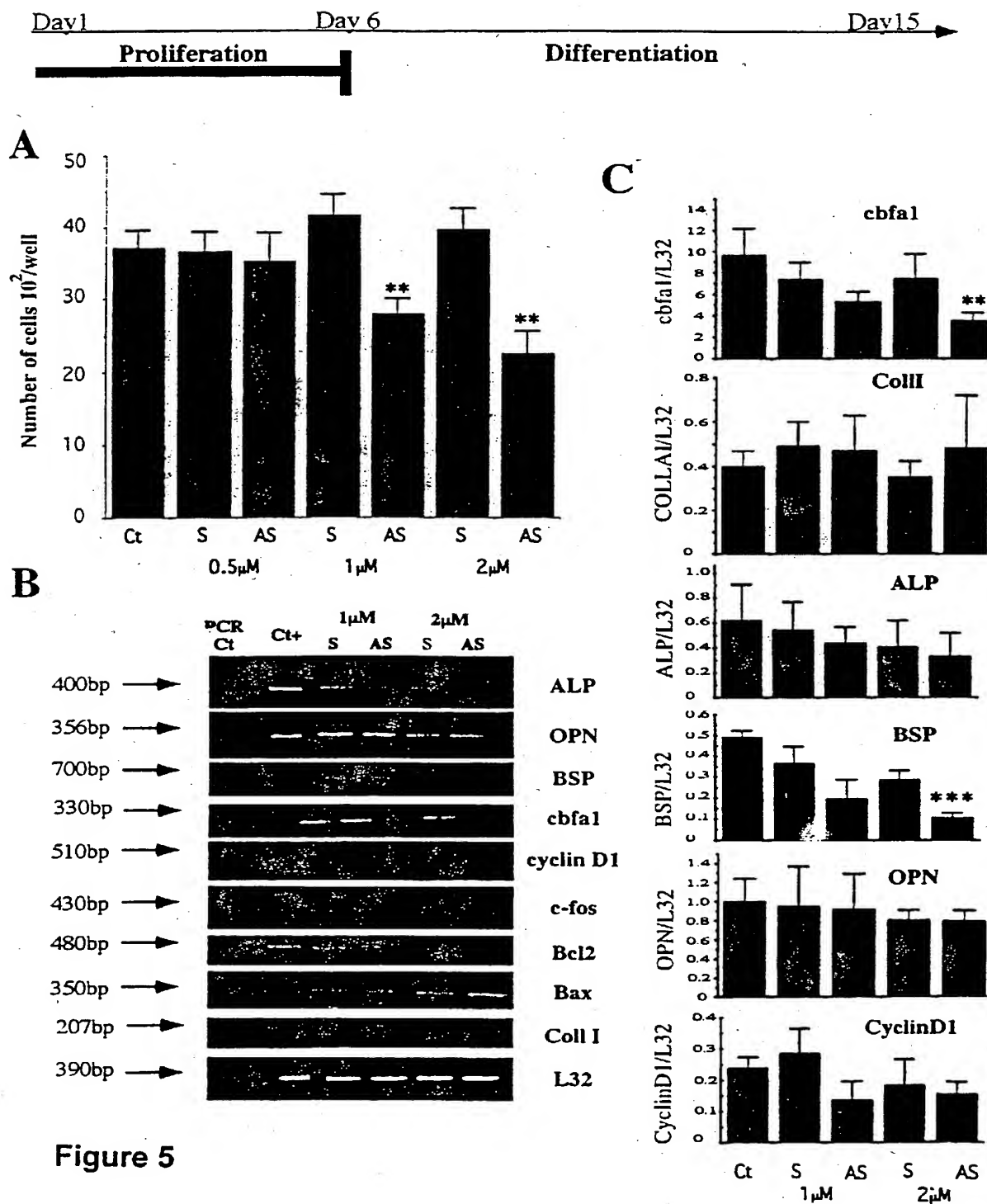


Figure 4

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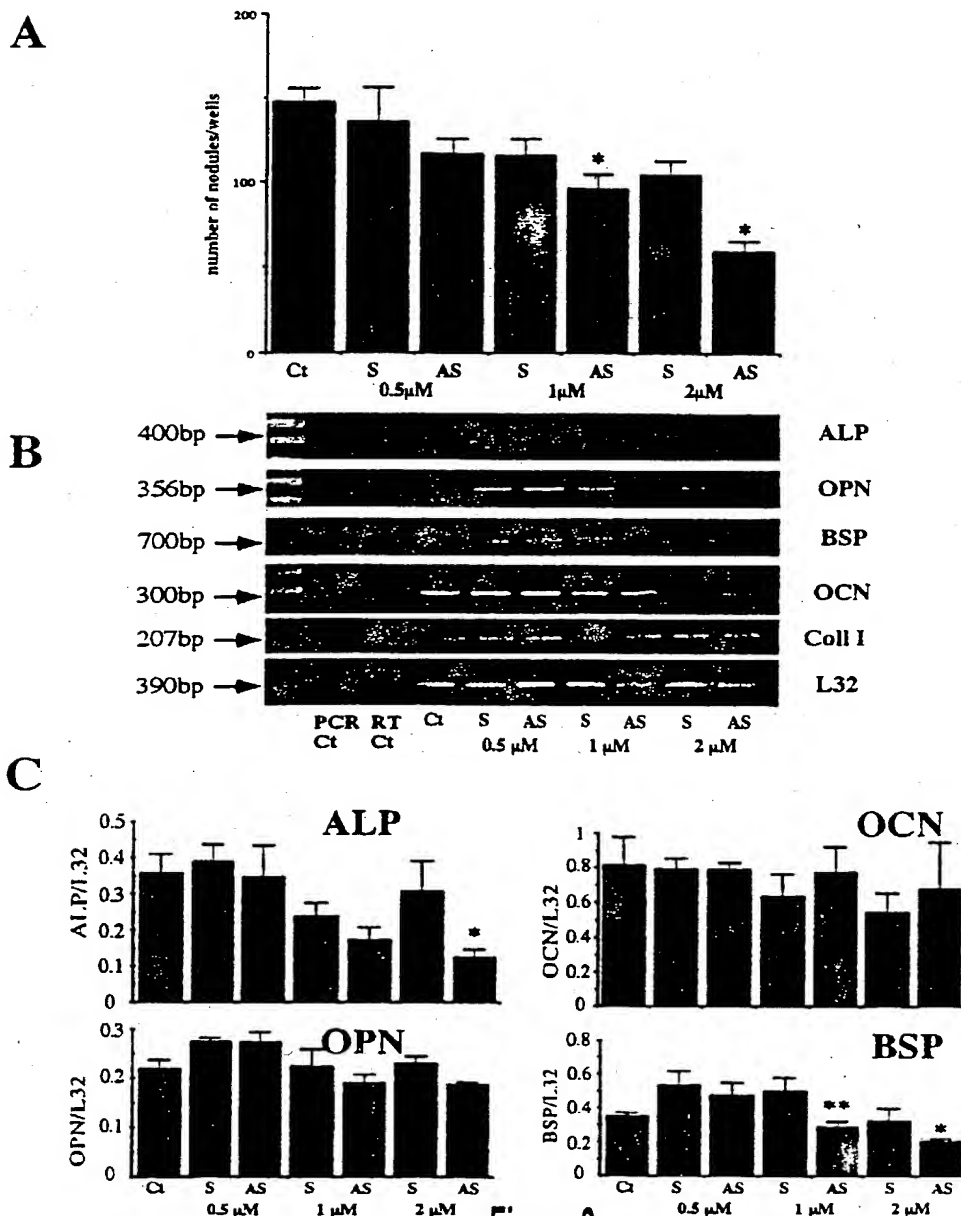
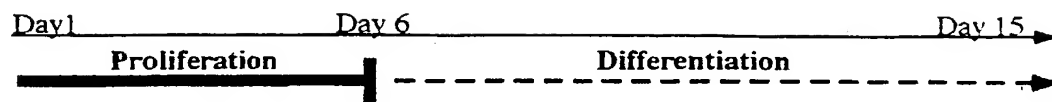


Figure 6

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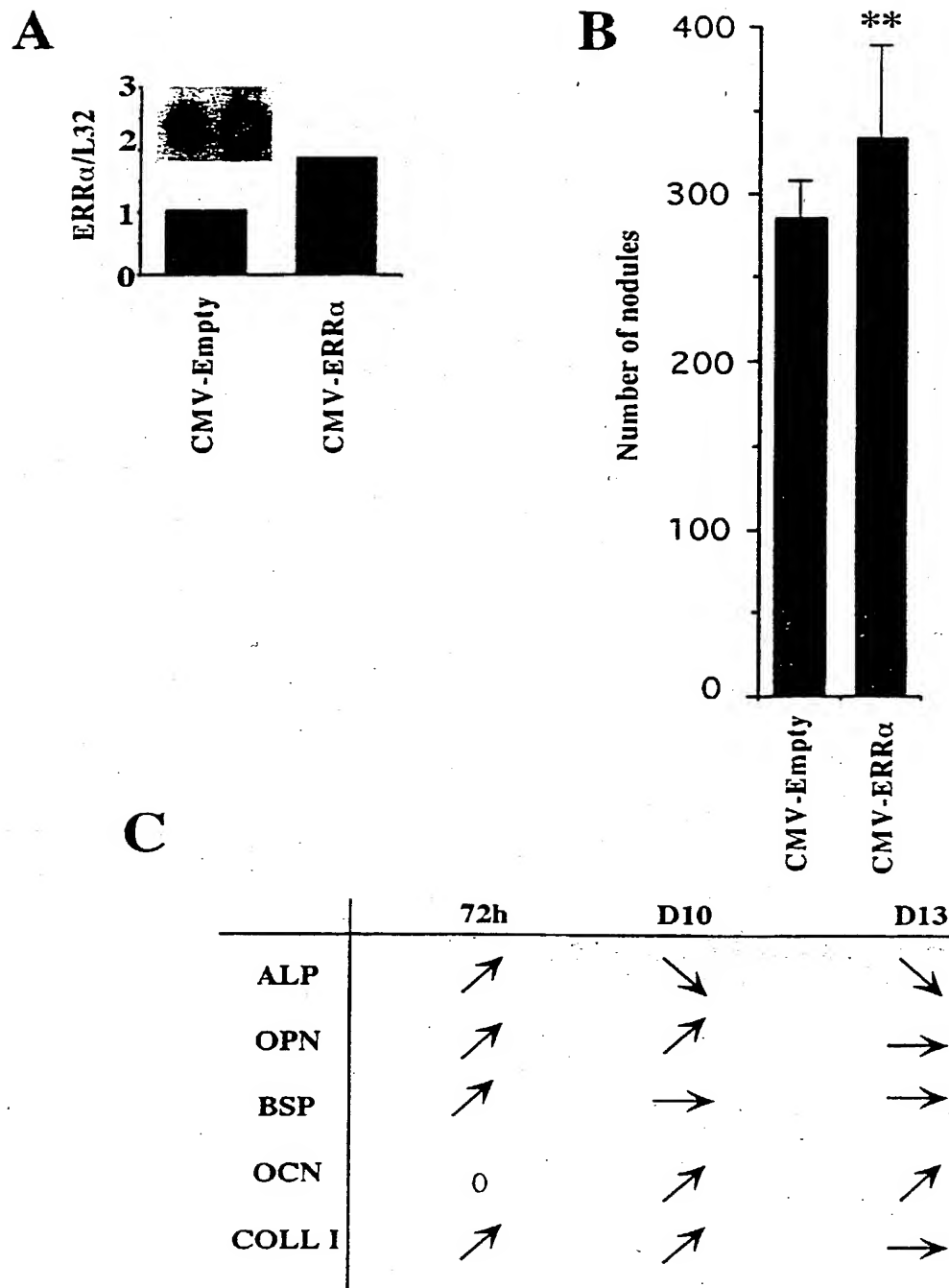
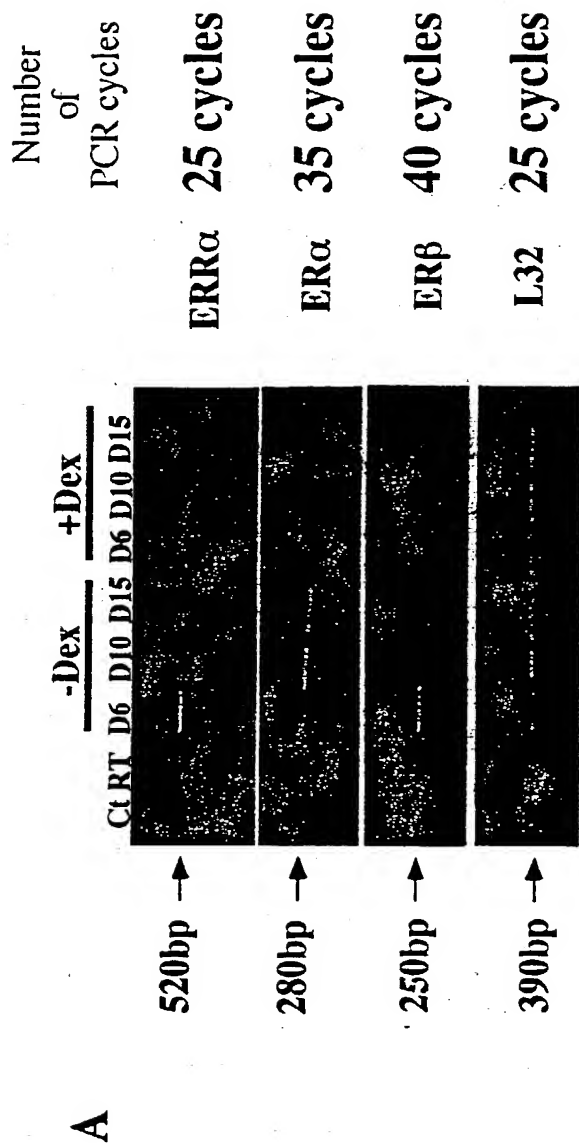


Figure 7



B

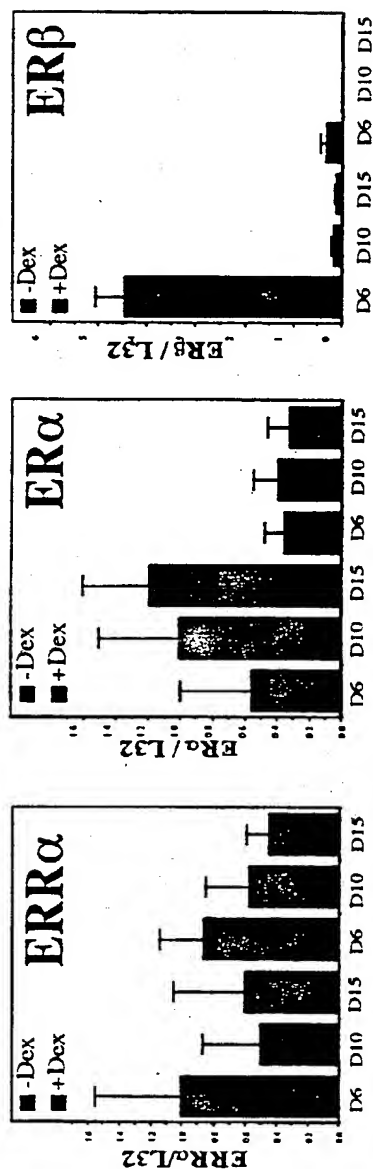


Figure 8

Culture time (days)

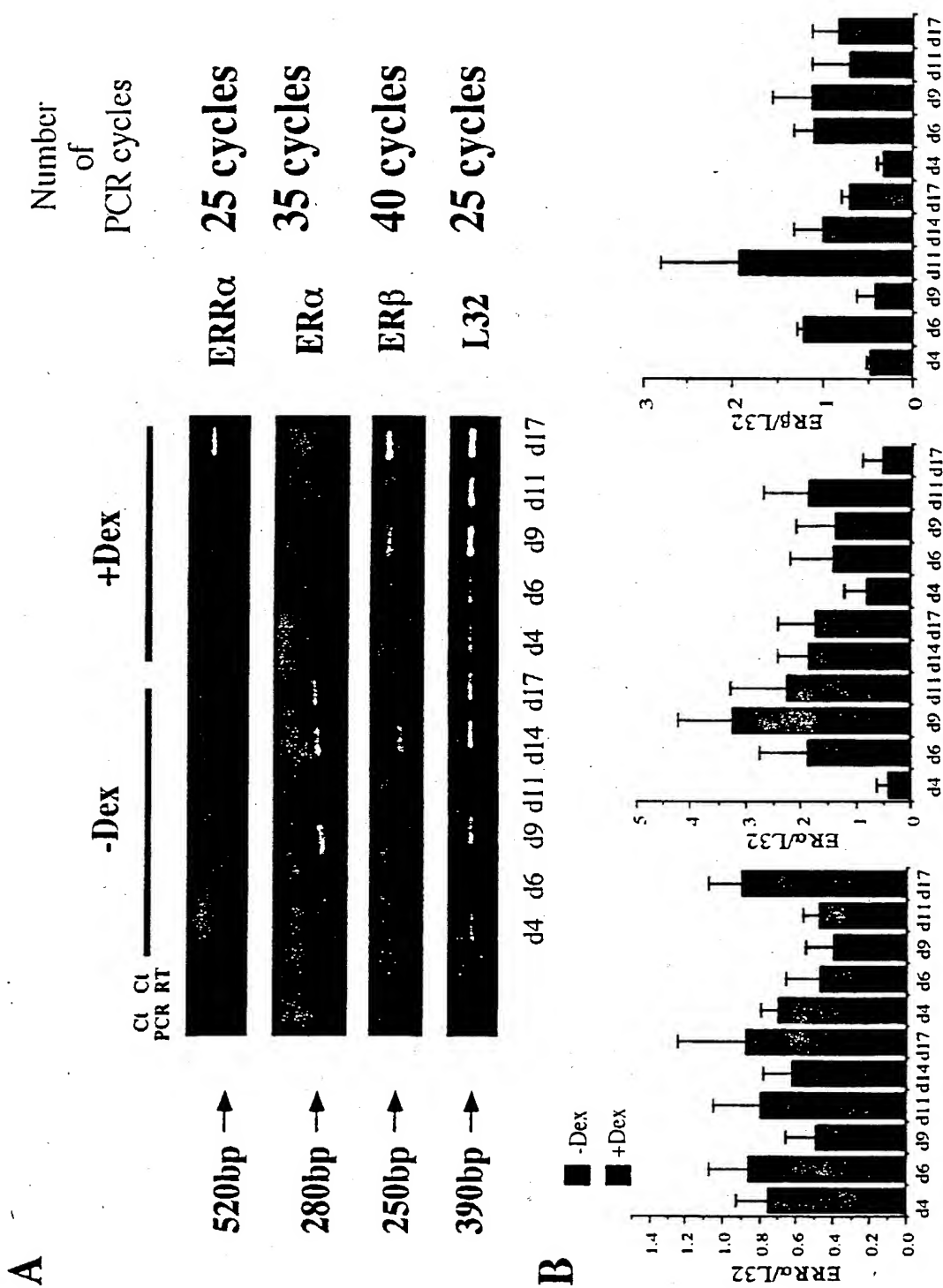
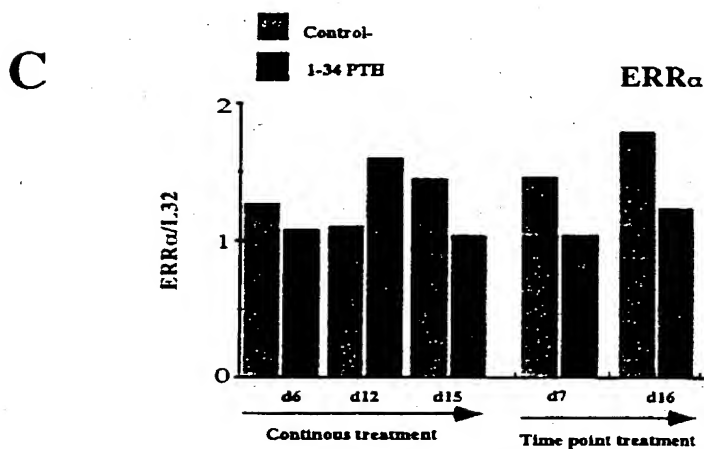
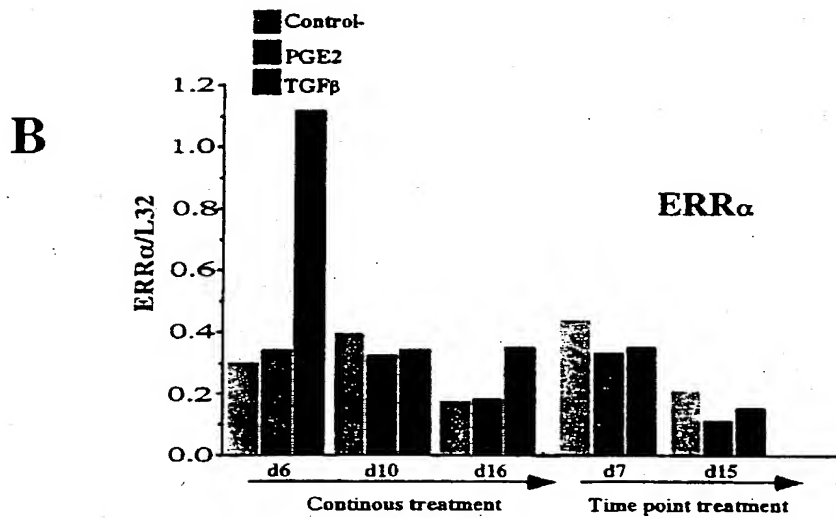
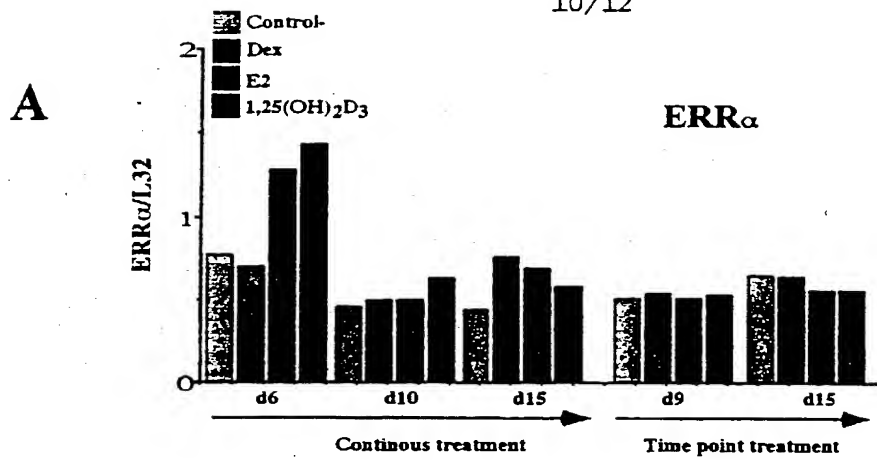


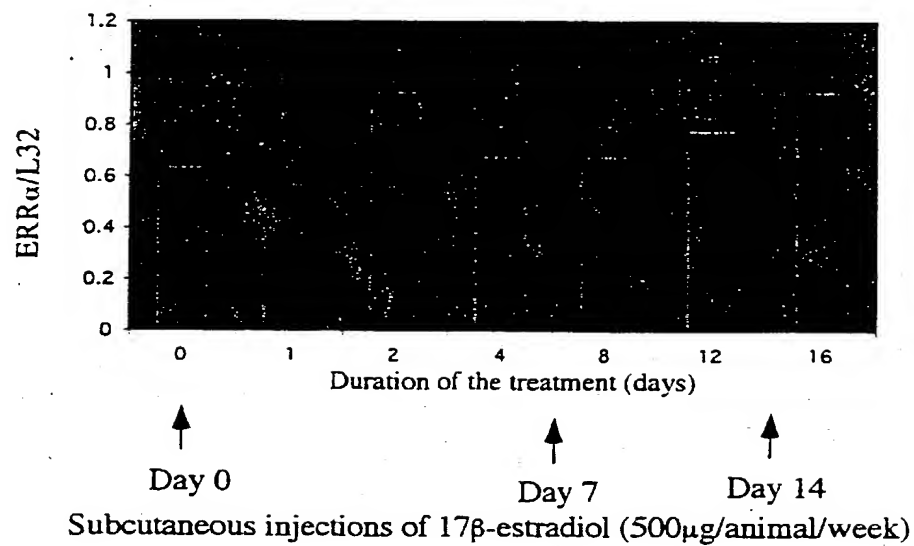
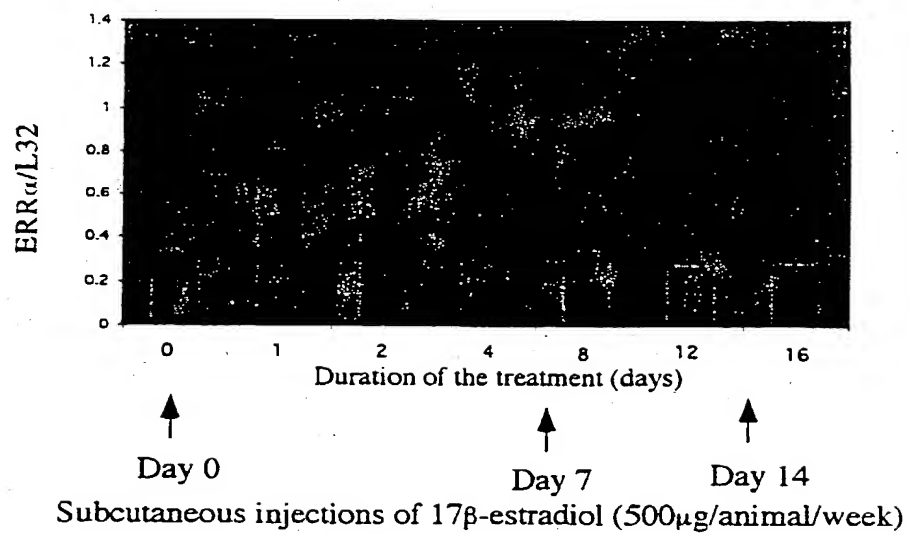
Figure 9

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A Femur**B** Flushed bones**Figure 11**

**Cortical
bone**

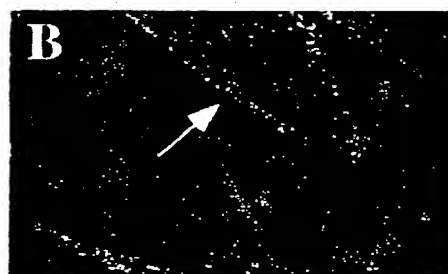
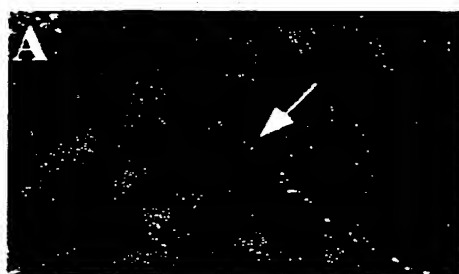
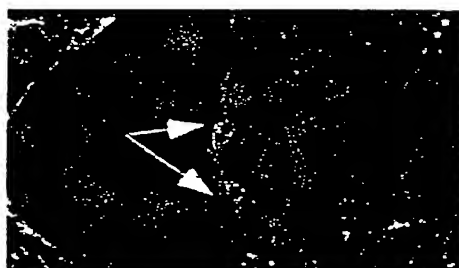


Figure 12



Calvaria



DECLARATION AND POWER OF ATTORNEY FOR PATENT APPLICATION

Attorney Docket No. 3477-95

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled **ESTROGEN RELATED RECEPTOR, ERR α , A REGULATOR OF BONE FORMATION**,

the specification of which

☐ is attached hereto

OR

☒ was filed on March 25, 2002 as United States Application No. 10/089,429 or PCT International Application Number _____ and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37 Code of Federal Regulations, §1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, § 119(a)-(4) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States of America, listed below and have also identified below any foreign application for patent or inventor's certificate, or of any PCT International application having a filing date before that of the application on which priority is claimed.

Number	Country	MM/DD/YYYY Filed	<input type="checkbox"/> Yes <input type="checkbox"/> No Priority Claimed
Number	Country	MM/DD/YYYY Filed	<input type="checkbox"/> Yes <input type="checkbox"/> No Priority Claimed
Number	Country	MM/DD/YYYY Filed	<input type="checkbox"/> Yes <input type="checkbox"/> No Priority Claimed

I hereby claim the benefit under Title 35, United States Code, § 119(c) of any United States provisional application(s) listed below.

2,284,103	09/30/1999
Application Number(s)	Filing Date (MM/DD/YYYY)

	Filing Date (MM/DD/YYYY)
Application Number(s)	

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) or § 365(c) of any PCT international application designating the United States of America, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application(s) in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application (37 C.F.R. § 1.63(d)).

Appn. Serial No.	Filing Date	Status Patented/Pending/Abandoned
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Appn. Serial No.	Filing Date	Status Patented/Pending/Abandoned
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Appn. Serial No.	Filing Date	Status Patented/Pending/Abandoned
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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Handwritten musical score for the song "The Rose Tree". The score is written on a single page of aged, yellowed paper. It features a single melodic line in a treble clef, with a key signature of one sharp (F#). The music is written in a cursive, handwritten style. The lyrics are written below the notes, and the title "The Rose Tree" is written at the top right. The score includes a variety of musical notations, including eighth, sixteenth, and thirty-second notes, as well as rests and bar lines. The paper shows signs of age, with some staining and discoloration.

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Karen A. Magni
Myers Bigel Sibley & Sajovec
Post Office Box 37428
Raleigh, NC 27627

(919)854-1400

(919) 854-1401

Jane R. Aubin

Date: 06 August 2002

643 Pape Avenue
Toronto, ON M5K 3S2
Canada

Canadian

Post Office Address:

1001 1001 001 0791739 5'2001

11032423 112302

Full name of second inventor: Edith Bonnellye ^{2nd}

Inventor's Signature: *Bonnellye* Date: 7/08/2002

Residence: 534 Huron Street
Toronto, ON M5R 2R7
Canada

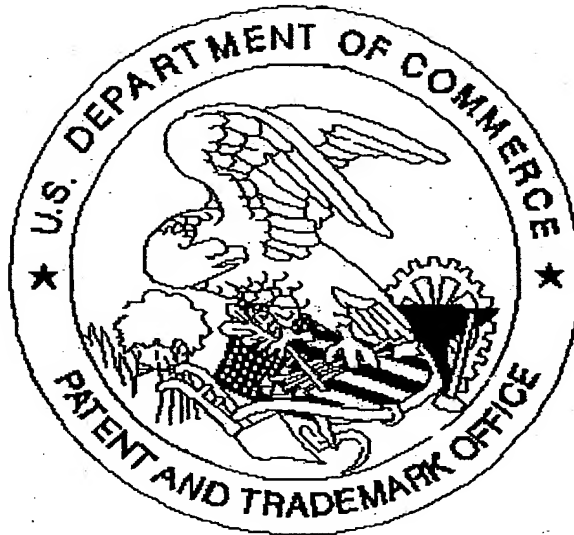
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